Protoscolicidal Effects of the Garlic Chloroformic Extract on the Protoscolices of Hydatid Cyst at a Short Exposure Time, up to Five Minutes

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Abstract

Background: The treatment of choice for hydatidosis as an important zoonotic disease is surgery. Different agents are injected into the cyst to prevent secondary hydatidosis. To avoid the side effects of such protoscolicidal agents, considering the high protoscolicidal effects of the garlic extract, we conducted the present study on protoscolices in limited applicable times and compared the extract with some chemical agents.

Methods: Sheep’s liver and lung cysts were collected. Ninety tubes were selected and divided into 3 sets (for different exposure times), each one comprising 5 groups of 6 tubes. Each tube contained 3000–4000 protoscolices. The groups were 0.5% cetrimide (as positive control), 20% hypertonic sodium chloride, 0.5% silver nitrate, 0.9% normal saline (as negative control), and the garlic chloroformic extract (200 mg/mL). The viability of the protoscolices was assessed using 0.1% eosin. The ANOVA and LSD were used to compare the mean viability of the protoscolices after exposure to the different agents at different times and concentrations. The data were analyzed using SPSS software, version 17. A P<0.05 was considered significant.

Results: Our findings showed that the protoscolicidal effects of the garlic extract at 1 (P<0.001) and 2 (P<0.001 and P=0.003) minutes of exposure were higher than those of sodium chloride and silver nitrate. At 5 minutes of exposure, there was no difference between the garlic extract and sodium chloride (P=0.36); however, the difference between these agents and silver nitrate was significant.

Conclusion: The garlic chloroformic extract in a short exposure time had high protoscolicidal effects and could substitute other agents.

What’s Known

- Different chemical agents are injected into the cyst before surgery to prevent secondary hydatidosis.
- Common protoscolicides often cause side effects such as liver necrosis and cholangitis.

What’s New

- Protoscolicidal effects of the garlic chloroformic extract within 30 minutes on protoscolices are comparable to those of cetrimide.
- Protoscolicidal effects of the garlic chloroformic extract in a limited exposure time (<5 min) on protoscolices can be applicable during hydatid cyst surgery.

Keywords ● Echinococcosis ● Garlic ● In Vitro techniques

Introduction

Hydatidosis/ echinococcosis is one of the most important zoonotic parasitic diseases, with a high parasite prevalence in many parts of the world, including Iran.¹-⁴ The causative agent is the larval stage of parasitic platyhelminth, Echinococcus granulosus (E. granulosus). Humans are infected by ingesting
parasite eggs released from infected dogs and canine species, and the infection is usually diagnosed via imaging and serologic tests.\textsuperscript{5-9} Consequently, in areas where \textit{E. granulosus} is endemic, care should be taken to prevent dogs from having access to raw offal and carcasses. Where dogs may have access to carcasses or raw viscera, especially from sheep, goats, cattle, or camels, they should be treated at least every 6 weeks with an effective anthelmintic containing praziquantel.\textsuperscript{1}

Cystic echinococcosis affects mainly the intermediate host’s internal organs such as the liver, lungs, and rarely spleen, kidneys, brain, and other organs.\textsuperscript{6,10} It has been suggested that surgery is still the most important treatment method with chemotherapy as the coadjuvant treatment; nevertheless, the leakage of protoscolices during surgery may cause secondary hydatidosis.\textsuperscript{11,12} The inoculation of protoscolicidal agents into the hydatid cyst in order to reduce the leakage risk of the viable protoscolices to the adjacent regions is a part of surgery or the PAIR technique (Puncture, Aspiration, Injection of scolicidal agent, Reaspiration).\textsuperscript{13,14} They are used for destroying the germinal layer and killing the viable protoscolices of the hydatid cyst.\textsuperscript{15} Different protoscolicidal agents such as hypertonic saline, silver nitrate, and cetrimide have been used to sterilize cysts.\textsuperscript{16} In vitro studies using glucose 50% have shown that the hypertonic glucose solution is highly effective in killing the protoscolices of \textit{E. granulosus}.\textsuperscript{17} Common protoscolicidal agents like 20% hypertonic sodium chloride, 0.5% silver nitrate, cetrimide, and formalin cause severe complications such as liver necrosis and cholangitis.\textsuperscript{18} Therefore, the World Health Organization’s working group on hydatidosis proposed an urgent need for new protoscolicidals which would be more effective with fewer complications.\textsuperscript{18} Subsequent research has suggested garlic (\textit{Allium sativum}), the antiparasitic effects of which have been evaluated for a number of parasites such as \textit{Hymenolepis nana}, \textit{Ascaris lumbricoides}, and \textit{Giardia intestinalis}.\textsuperscript{19,20} Many in vitro studies on the protoscolicidal effects of herbal extracts have applied long exposure times between protoscolices and herbal extracts. A previously conducted investigation has shown the successful in vitro protoscolicidal effects of the chloroformic extract of garlic at 30 minutes of exposure or longer.\textsuperscript{11} However, such long exposure times are not applicable during surgery or the PAIR technique and shorter exposure times are needed before cyst evacuation. Accordingly, the present study was designed with exposure times of under 5 minutes (i.e., 1 min, 2 min, and 5 min) in order to assess the applicability of the chloroformic extract of garlic as a usable protoscolicidal agent during surgery or the PAIR technique. A number of other protoscolicidal agents such as cetrimide were also used for the comparison of the protoscolicidal effects of the garlic chloroformic extract.

### Materials and Methods

The agents used in the present study were 0.5% cetrimide, 20% sodium chloride, 0.5% silver nitrate, normal saline, and a solution of the garlic chloroformic extract (200 mg/mL). The garlic chloroformic extract was prepared as follows: 150 g of fresh garlic (Hamadan origin) was crushed and placed in a succilate apparatus, and while it was being warmed for 24 hours, 400 mL of chloroform was added. The extract was desiccated with a rotary evaporator to remove the extra chloroform solvent, and the remaining powder in the form of crystal was used for the assay. A negative control group was also considered using 0.9% normal saline. Hydatid cysts of sheep’s liver and lungs were carried to the laboratory, and protoscolices measuring a total of 30000/mL to 40000/mL were collected in a sterile condition. The samples of protoscolices were selected while the viability was more than 90%. A total of 90 tubes were selected and divided into 3 sets each with 5 groups of 6 tubes. Each set was used for different exposure times of 1, 2, and 5 minutes. Each tube was transferred to 0.1 mL of a fluid containing 3000–4000 protoscolices in a sterile condition. The groups were divided as follows: (1) 0.5% cetrimide, (2) 0.5% silver nitrate, (3) 20% hypertonic sodium chloride, (4) garlic chloroformic extract (200 mg/mL), and (5) normal saline as the negative control. The tests were carried out at exposure times of 1, 2, and 5 minutes in each group, and the viability of the protoscolices was assessed using 0.1% eosin.

The experimental protocols were approved by the Ethics Committee of Shiraz University of Medical Sciences.

After 1 minute of exposure, 10 mL of normal saline was added. The upper fluid (supernatant) was discarded after centrifugation at 500 rpm and the procedure was repeated 3 times. The viability of the protoscolices was evaluated using 0.1% eosin.\textsuperscript{17} For the validity test, 0.01 mL of the exposed protoscolices was transferred onto a slide and mixed with 0.01 mL of 0.1% eosin to look for dead or living protoscolices.\textsuperscript{21} The protoscolicidal ability of each agent was repeated 6 times. At least 250 protoscolices
were counted in each sample while the dead and living protoscolices were measured. The dead protoscolices were stained (figure 1A), while the living protoscolices remained colorless (figure 1B). These steps were also repeated at the exposure times of 2 and 5 minutes. The ANOVA test was utilized to compare the mean viability for the different materials. Additionally, the mean viability was compared between the group of agents via the least significant difference (LSD) method. The data were analyzed using SPSS software, version 8. A P<0.05 was considered statistically significant. The mean percentage of the dead protoscolices and the agents used for each time of exposure are summarized in tables 1 to 3.

**Results**

The exposure results of the protoscolicidal agents are shown in tables 1, 2, and 3. In vitro observations at 5 minutes of exposure showed no difference between the garlic chloroformic extract (200 mg/mL) and 20% sodium chloride (P=0.36); however, the difference between these agents and 0.5% silver nitrate was significant (P<0.001) (table 3).

According to table 1, while the protoscolicidal effects of the garlic chloroformic extract (200 mg/mL) at 1 minute were higher than those of 0.5% silver nitrate and 20% sodium chloride (P<0.001), the effects were similar to those of 0.5% cetrimide (P=0.36).

At 1 minute of exposure, the protoscolicidal effects of the garlic chloroformic extract (200 mg/mL) were significantly different from those of 0.5% silver nitrate and 20% hypertonic sodium chloride (P<0.001 and P=0.003).

The application of the agents at 2 minutes of exposure revealed no statistically significant difference in terms of protoscolicidal effects between the garlic chloroformic extract (200 mg/mL) and 0.5% cetrimide (table 2). Nonetheless, the ANOVA test demonstrated statistically significant differences regarding protoscolicidal effects between 0.5% cetrimide and the other agents (P<0.001). Table 2 shows that while the mean protoscolicidal effect of the garlic chloroformic extract (200 mg/mL) was comparable with that of 0.5% cetrimide, it was statistically different from that of the 20% sodium chloride, 0.5% silver nitrate, and normal saline groups (P<0.05) (figure 1A and figure 1B).

**Discussion**

Different protoscolicidal agents have been used for hydatidosis as an important zoonotic disease. Nevertheless, the long exposure

<table>
<thead>
<tr>
<th>Agents Groups</th>
<th>Test reagents</th>
<th>n</th>
<th>Mean protoscolices exposed</th>
<th>Mean protoscolices dead after Exposure±SD</th>
<th>% dead/total</th>
<th>LSD* P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Cetrimide 0.5%</td>
<td>6</td>
<td>255</td>
<td>255±0</td>
<td>100%</td>
<td>1 vs. 2: &lt;0.001</td>
</tr>
<tr>
<td>2</td>
<td>0.5% Silver nitrate</td>
<td>6</td>
<td>261</td>
<td>148±1.5</td>
<td>56.7%</td>
<td>2 vs. 1: &lt;0.001</td>
</tr>
<tr>
<td>3</td>
<td>20% Hypertonic sodium chloride</td>
<td>6</td>
<td>255</td>
<td>225±1.6</td>
<td>88.2%</td>
<td>3 vs. 1: &lt;0.001</td>
</tr>
<tr>
<td>4</td>
<td>Garlic chloroformic extract (200 mg/mL)</td>
<td>6</td>
<td>253</td>
<td>251±0.5</td>
<td>99.2%</td>
<td>4 vs. 1: 0.037</td>
</tr>
<tr>
<td>5</td>
<td>Normal saline</td>
<td>6</td>
<td>265</td>
<td>5±0.6</td>
<td>1.8%</td>
<td>5 vs. 1: &lt;0.001</td>
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*Least significant difference; **P value for one-way ANOVA was <0.001
time of their application is not suitable for hydatid cyst surgery or the PAIR technique. The World Health Organization has suggested that the protoscolicidal effects of protoscolicidal agents be compared to those of cetrimide. In the present study, this agent showed a 100% protoscolicidal effect at 1 minute of exposure. In addition, the protoscolicidal effects of the garlic chloroformic extract at 1 minute of exposure were similar to those of cetrimide, although the protoscolicidal effects of these 2 agents had a significant difference with those of the other agents used. Silver nitrate solution has been used since 1963. It has been observed that its 0.5% solution is more effective than its 1% solution. Protoscolices are killed within 30 seconds when exposed to 0.5% silver nitrate; their internal structure is separated and they form an individual granule. Be that as it may, in the present study, 0.5% silver nitrate exhibited only a 94% protoscolicidal effect. A concentrated salty

<table>
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<th>Table 2: Protoscolicidal effects of the garlic chloroformic extract in comparison with cetrimide, silver nitrate, and sodium chloride solutions at 2 minutes of exposure **</th>
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<tr>
<td><strong>Agents Groups</strong></td>
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<tr>
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**P value for one-way ANOVA was<0.001**

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<th>Table 3: Protoscolicidal effects of the garlic chloroformic extract in comparison with cetrimide, silver nitrate, and sodium chloride solutions at 5 minutes of exposure**</th>
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<td><strong>Agents Groups</strong></td>
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**P value for one-way ANOVA was <0.001**
solution will cause the lysis and devastation of protoscolices by establishing a proper osmotic difference in the cuticle or outer layer. A number of studies have demonstrated that sodium chloride at concentrations of 3% and 10% has no protoscolicidal effect, while at a 20% concentration and at 15 minutes of exposure, it is capable of devastating all protoscolices. In the present study, this agent showed 89%, 94.8%, and 99.6% protoscolicidal effects at exposure times of 1, 2, and 5 minutes, respectively.

Several studies have investigated the protoscolicidal effects of different anthelmintic drugs. For example, in an in vitro study, the effects of flubendazole at concentrations of 1, 5, and 10 µg/mL on the protoscolices of *E. granulosus* were examined and the results showed that the first signs of protoscolices destruction appeared 3 days after exposure to this drug; still, its obvious protoscolicidal effect (i.e., decrease in protoscolex viability) was seen 18 days after the drug exposure. Additionally, 25 days after the exposure of the protoscolices to fluconazole at a concentration of 5 µg/mL, only 13.9% of the protoscolices were viable and protoscolex mortality after 30 days of exposure was 100%. In that study, the validity test results were indicative of tissue damage at the ultrastructural level. Hosseini et al. compared protoscolicidal effects between hypertonic glucose at concentrations of 10, 15, 25, and 50% and some protoscolidal agents such as 0.5% cetrimide (as a positive control), 0.5% silver nitrate, 20% hypertonic saline, and normal saline (as a negative control) at the exposure times of 1, 2, and 5 minutes and reported that the high concentration of glucose (50%) had a stronger protoscolicidal effect than 0.5% silver nitrate but this effect was similar to that of 20% hypertonic saline.

The importance of protoscolicidal agents is still considerable in the treatment of hydatid disease; nonetheless, the ideal substances are those which are highly effective, inexpensive, and quick-acting with the fewest complications. Since many of the protoscolidal agents are not safe enough and induce some side effects, there is a tendency to use natural substances as protoscolidal solutions. In an in vitro investigation, the essential oils of *Mentha piperita* and *Mentha pulegium* were used separately and both together as protoscolidal agents at different concentrations and exposure times on the protoscolices of *E. granulosus*. The results indicated that *Mentha pulegium* had an obviously more potent protoscolicidal effect, with its maximum effect seen 12 days after incubation. Additionally, transmission electron microscopy proved the ultrastructural damage to the protoscolices caused by this essential oil. In another in vitro study, the protoscolicidal effects of the hydroalcoholic extract of *Satureja khuzestanica* leaves and the hydrated extract of *Olea europaea* leaves were examined at different concentrations and exposure times. Among them, concentrations of 0.1% and 0.01% had strong protoscolicidal effects at the exposure time of 120 minutes. *Satureja khuzestanica* at a concentration of 0.1% had a very strong protoscolicidal effect at the exposure times of 30, 60, and 120 minutes.

Garlic has a special position among herbal and traditional medicines and its antibacterial, antifungal, and antiparasitic effects have been confirmed in many studies. The scolicidal effects of *Allium sativum* flowers on hydatid cyst protoscolices have also been used with success. Various extracts of garlic such as hydrated, alcoholic, and chloroformic extracts have exhibited significant effects on parasites such as *Hymenolepis nana*, *Leishmania*, and *Giardia intestinalis*. Their long-term (>30 min) protoscolicidal effects were confirmed in an in vitro study, which showed that the garlic chloroformic extract at a concentration of 200 mg/mL exerted good protoscolicidal effects on the protoscolices of hydatid cysts at an exposure time of 1 hour and longer. However, such long exposure times are not applicable during hydatid cyst surgery. We used the garlic chloroformic extract in the current study at exposure times much shorter than those in previous research.

In another in vitro study, the protoscolicidal effects of the metanolic extract of *Allium sativum* at 2 concentrations of 25 and 50 mg/mL at the exposure times of 10, 20, 30, 40, and 60 minutes were evaluated and the results showed that the protoscolical effect of the 50 mg/mL concentration was 100% after 10 minutes of exposure. In contrast, in the present study, the garlic chloroformic extract (200 mg/mL) had a 100% protoscolicial effect at shorter exposure times of 2 and 5 minutes. Among the agents tested, the garlic chloroformic extract, which has previously been evaluated in animals with acceptable results, can be introduced as a new and desirable protoscolidal agent.

**Conclusion**

According to the results of the present study, the chloroformic extract of garlic (200 mg/mL) has a high protoscolicial effect at a short exposure time and it could, therefore, substitute the protoscolidal agents in current use. To determine both whether the garlic extract has
any side effects in the human body and what concentrations and exposure times are suitable for cyst surgery or the PAIR technique, we recommend ex vivo and in vivo studies on this issue.

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Conflict of Interest: None declared.

References


