

Effect of Alpha-1 Antitrypsin and Irisin on Post-Exercise Inflammatory Response: A Narrative Review

Marta Pawłowska, PhD;  Celestyna Mila-Kierzenkowska, PhD

Department of Medical Biology and Biochemistry, Ludwik Rydygier Collegium Medicum in Bydgoszcz, Nicolaus Copernicus University in Toruń, Poland

Correspondence:

Marta Pawłowska, PhD;
Department of Medical Biology and Biochemistry, Ludwik Rydygier Collegium Medicum in Bydgoszcz, Karłowicza 24, Postal code: 85-092, Bydgoszcz, Poland
Tel: +48 52 5853822

Email: marta.pawlowska@cm.umk.pl

Received: 13 December 2022

Revised: 12 January 2023

Accepted: 16 February 2023

Abstract

Physical activity has a positive effect on human health and emotional well-being. However, in both amateur and professional athletes, training poses a risk of acute or chronic injury through repetitive overloading of bones, joints, and muscles. Inflammation can be an adverse effect of intense exercise caused by several factors including oxidative stress. The present narrative review summarizes current knowledge on inflammatory markers induced by physical exercise. Post-exercise recovery may reduce inflammatory responses and is key to effective training and adaptation of muscle tissues to sustained physical exertion.

Please cite this article as: Pawłowska M, Mila-Kierzenkowska C. Effect of Alpha-1 Antitrypsin and Irisin on Post-Exercise Inflammatory Response: A Narrative Review. *Iran J Med Sci.* 2024;49(4):205-218. doi: 10.30476/IJMS.2023.97480.2925.

Keywords • Exercise • Inflammation • Serine proteinase inhibitors • Oxidative stress

What's Known

- Activation of inflammatory response is essential for muscle repair and adaptation to exercise. However, excessive or persistent inflammation can lead to further tissue damage due to the nonspecific phagocytic function of inflammatory cells.
- Reducing inflammation has a beneficial effect on the recovery of muscle function after intense exercise.

What's New

- Alpha-1 antitrypsin is a serine protease inhibitor involved in inflammatory responses.
- Blood biomarkers provide exact and reliable data to assess muscle damage and exercise-induced inflammation. These markers offer valuable insight into the relative state of recovery. Post-exercise recovery is key to effective training and adaptation to sustained physical exertion.

Introduction

In recent years, there has been a growing awareness of the need for a healthy lifestyle, especially regular exercise. It is known that physical activity has a positive effect on overall health and can reduce the risk of severe health problems such as obesity and cardiovascular diseases.^{1, 2} However, a vigorous workout is often associated with the development of microdamage in muscle tissue, which can lead to muscle soreness.³ Functional recovery of muscles after tissue microdamage may take up to several days.⁴ As a post-exercise response, intense physical activity may induce inflammation.² Therefore, the main challenge in sports medicine is to develop more effective methods to regenerate muscle tissue and accelerate the post-exercise recovery process. In particular, the type of recovery, and its ability to reduce the inflammatory response seems to be crucial for effective training and adaptation of muscle tissue to sustained physical exertion. The present narrative review summarizes current knowledge on exercise-induced inflammatory markers, and the effect of alpha-1 antitrypsin (AAT) and irisin on post-exercise inflammatory response. We also discuss the role of oxidative stress in triggering inflammation after physical activity. The findings of this study will help sports medicine professionals develop appropriate countermeasures against the adverse health effects of intense exercise.

Course of the Inflammatory Process

Inflammation is a defense system in response to disrupted

homeostasis.^{5,6} Tissue damage is known to trigger signals that activate the host's immune system against infectious agents to promote tissue repair.⁷ Inflammatory response affects tissue homeostasis, increases blood flow, facilitates immune cell activation and migration, and triggers secretion of cytokines, chemokines, and growth factors.^{8,9} Acute inflammation ultimately leads to the elimination of infectious agents, of damaged tissues, clearance of inflammatory cells, and return to homeostasis.¹⁰ Persistent acute inflammation may develop into chronic inflammation, which can cause permanent tissue damage.¹¹ The process of inflammation is initially targeted locally and then centrally by inflammatory mediators circulating in the blood.¹² Neutrophils are key mediators and the first immune cells to respond to inflammation. These cells regulate acute inflammatory response by rapid release of reactive oxygen species (ROS), i.e., oxidative burst.¹³ Neutrophils migrate to the site of injury through the bloodstream as a result of vasodilation and can increase vascular permeability caused by basophils or mast cell degranulation, activation of the complement system, or release of prostaglandins and leukotrienes.¹³ In the early stage of endothelial injury, leukocytes migrate to the injury site for endothelial adhesion to the tissue.¹⁴ Activation of neutrophils results in the release of many chemicals capable of attacking pathogens and mediators to activate other cells such as monocytes. Activated monocytes, in turn, produce and secrete interleukins, particularly interleukin 6 (IL-6), and other cytokines.⁵ These mediators cause a nonspecific inflammatory response by inducing the production of acute-phase proteins (APPs), primarily by the liver, such as C-reactive protein (CRP), serum amyloid A (SAA), fibrinogen, ferritin, AAT, ceruloplasmin, components of the complement system, and coagulation factors.¹⁵ APPs affect homeostasis and thus trigger defensive and adaptive processes that contribute to short-term healing. However, they can also lead to chronic inflammation and tissue damage.¹³ Next to macrophages and neutrophils, mast cells are also activated during the inflammatory response. These cells release histamine, proteolytic enzymes, and chemokines that significantly contribute to the progression of inflammation.¹⁶

In summary, inflammation is the response of the immune system to harmful stimuli, such as cell damage. The inflammatory response is a defense mechanism essential to maintaining homeostasis in organisms by suppressing noxious stimuli and initiating the healing process. Cellular and molecular interactions minimize

injury during acute inflammatory responses. However, uncontrolled acute inflammation can become chronic and contribute to various inflammatory diseases.

Inflammatory Markers

Activation of neutrophils releases inflammatory mediators that regulate the inflammatory cascade.¹⁷ Secretion of chemokines and cytokines plays a significant role in modulating the inflammatory response through their capacity for cell surface binding and cell activation.¹⁶ Cytokines are small secreted proteins synthesized by multiple cells in response to various stimuli.¹⁸ They can be classified as pro-inflammatory, anti-inflammatory, and multifunctional cytokines.^{16,19} They are pleiotropic because they affect different types of cells, and their effect depends on the type of cell they target.²⁰ Cytokines bind to specific receptors on cells with different origins and functions. In addition, they mediate signal transduction via various intracellular messengers and transcription factors.¹⁸ Table 1 presents a list of cytokines and myokines that play a key role in the development of inflammatory processes.

The short half-life of cytokines suggests that these soluble mediators are rapidly eliminated under certain physiological conditions, thus ensuring their limited bioactivity.¹³ Moreover, cytokines can modulate the response of an organism to stimuli by activating the hypothalamic-pituitary-adrenal axis.²⁶ The final stage of acute inflammatory response is dominated by mediators involved in suppressing inflammation,²⁷ during which they control the spread of inflammation, limit inflammatory responses, and facilitate the final stage of muscle repair.¹⁶

Characteristics of Cytokine Response to Exercise

IL-6 is a multifunctional cytokine that plays an important role in activating host defense mechanisms by regulating the immune response, mediating acute phase response, and hematopoiesis.^{28,29} It promotes B-cell differentiation,²⁹ an important inducer of APPs in liver cells, and is involved in the proliferation and differentiation of T cells.^{28,30} Therefore, it might be a valuable marker of stress and muscle recovery.³¹ IL-6 also increases adipose tissue lipolysis, improves insulin sensitivity,³² and induces an increase in the production of anti-inflammatory cytokines such as IL-10.³³ IL-6 is an important marker of inflammation, because an increase in its concentration is associated with an increase in APPs concentration.³⁴

Table 1: Cytokines and myokines that play a key role in the development of inflammatory response

Cytokine/Myokine	Origin	Function	Article
Interleukin 1 (IL-1)	Macrophages, dendritic cells, T and B lymphocytes	Activation of T and B lymphocytes, enhancing the production of other cytokines and acute phase proteins, induction of adhesive molecules	Germolec et al. ¹³
Interleukin 6 (IL-6)	Macrophages, dendritic cells, B cells, activated T cells	Development of myeloid cells, regulation of acute phase proteins	Germolec et al. ¹³
Interleukin 8 (IL 8)	Monocytes, macrophages	Recruitment and activation of immunocompetent cells, stimulation of chemotaxis, and angiogenesis	Remick ²¹
Interleukin 10 (IL-10)	Macrophages, dendritic cells, regulatory B and T lymphocytes	Inhibition of pro-inflammatory cytokines	Germolec et al. ¹³
Transforming growth factor beta (TGF- β)	Macrophages, megakaryocytes, chondrocytes	Inhibition of cytokine production and activity, inhibition of B-cell proliferation, stimulation of wound healing	Germolec et al. ¹³
Tumor necrosis factor alpha (TNF- α)	Macrophages, dendritic cells, lymphocytes, mast cells	Increase in MHC expression, macrophage activation, enhancing the destruction of cancer cells	Germolec et al. ¹³
Irisin	Muscle cells	Inhibition of pro-inflammatory cytokines	Lin et al. ²²
Decorin	Muscle cells, fibroblasts, endothelial cells	Cell-matrix crosstalk modulation, promotion of hypertrophy of muscle fibers	Sabouri et al. ²³
Apelin	Adipocytes, monocytes	Regulation of energy homeostasis	Ramezani et al. ²⁴
Musclin	Muscle cells	Regulation of energy homeostasis	Clark et al. ²⁵

Tumor necrosis factor alpha (TNF- α) is a polypeptide cytokine produced by monocytes and macrophages that acts as a multipotent modulator of the immune response.³⁵ TNF- α in the circulation activates neutrophils, changes the properties of vascular endothelial cells, and regulates the metabolic activity of tissue in response to infection or tissue damage.³⁵ Transforming growth factor beta (TGF- β) is another pleiotropic cytokine with strong immunoregulatory properties. TGF- β family includes six isoforms, but only TGF- β 1, TGF- β 2, and TGF- β 3 have been identified and expressed in mammals. TGF- β regulates fundamental aspects of cellular function, such as growth, differentiation, adhesion, migration, apoptosis, extracellular matrix production, and inflammation.³⁶

Interleukin 8 (IL-8) is a pro-inflammatory chemokine secreted by monocytes, macrophages, and endothelial cells that activates neutrophils.³⁷ It plays an important role in regulating the acute inflammatory response. It is quickly synthesized at the site of inflammation, where it recruits and activates immunocompetent cells.²³ IL-8 is not only released early in the inflammatory response, but its levels remain increased for several days or weeks.²³ Another pro-inflammatory cytokine is interleukin-1 beta (IL-1 β), a member of the interleukin-1 family, which can stimulate T and B lymphocytes, increase cell proliferation, and initiate or suppress gene expression of several proteins.³⁸ Whereas interleukin 10 (IL-10) is an anti-inflammatory cytokine that plays a vital

role in regulating the functions of lymphoid and myeloid cells. It also regulates the proliferation and differentiation of B cells, mast cells, and thymocytes.³⁹

Characteristics of Lysosomal Enzyme Response to Exercise

Lysosomes are organelles involved in the intracellular digestion of various macromolecules that have entered the cell through endocytosis, and those derived from the cell's cytoplasm.⁴⁰ They are also involved in the apoptosis process.⁴¹ Lysosomal enzymes include about 40 acid hydrolases, namely lipases and phospholipases, nucleases, phosphatases (e.g., acid phosphatase), sulfatases (e.g., arylsulfatase A), proteases (e.g., cathepsin D), and other glycosidases.⁴²

Acid phosphatase (ACP) is a lysosomal enzyme associated with cells' physiological and pathological functions⁴³ and is present in different cells and tissues. In particular, epithelial cells of the prostate gland are characterized by high ACP activity.⁴⁴ It catalyzes the hydrolysis of phosphate monoesters in acidic conditions (pH 4-7).⁴⁵ In addition, it is an enzyme involved in the degradation of phagocytosed materials found in neutrophils and monocytes.⁴⁶ ACP also plays a role in the biological activation of lymphocytes, their blastic transformation, and the removal of cell organelles during mitosis.⁴⁷

Arylsulfatase A (ASA) belongs to the group of sulfatases that plays a role in many physiological processes.⁴⁸ Arylsulfatases can be expressed in tissues of various organs (e.g., liver, skin, lymph

nodes) and nervous tissue.⁴⁸ They can also be detected in urine and blood serum.⁴⁹ However, the highest activity of ASA is found inside lysosomes.⁴⁹ Human arylsulfatase catalyzes reactions at both acidic and neutral pH.⁵⁰ ASA catalyzes the degradation of sulfatides to galactosylceramides (GalC), starting with the hydrolysis of the sulfate residue.⁴⁸ Cathepsin D (CTSD) is a soluble lysosomal aspartic endopeptidase synthesized in the rough endoplasmic reticulum as preprocathepsin D.⁵¹ After removal of signal peptide, procathepsin D is directed to intracellular vesicular structures (lysosomes, endosomes, and phagosomes). In the cell, CTSD is localized in lysosomes and binds to the membranes of erythrocytes and macrophage endosome vesicles.⁵² The primary function of enzymatically active CTSD is the degradation of proteins in the acidic environment of lysosomes. Active forms of CTSD and other proteases can be released from lysosomes into the cytoplasm in response to apoptotic stimuli, thus contributing to cell death.^{51, 52} In addition, it participates in blood clotting and fibrinolysis.⁵¹ CTSD activates other proteases, such as cathepsin B, cathepsin L, collagenases, caspases-3, and caspases-9.⁵³ The proteolytic activity of CTSD is modulated by several physical and chemical factors, including pH, metabolic products, growth factors, exogenous and endogenous inhibitors, and hormones.⁵⁴

Characteristics of Alpha-1 Antitrypsin (AAT)

AAT is a protein belonging to the family of serine protease inhibitors, called serpins.⁵⁵ It is produced in various tissues and cells, including monocytes, pulmonary macrophages, and phagocytes; but primarily by hepatocytes.⁵⁶ AAT is an APP with anti-inflammatory properties.⁵⁷ Its main biochemical activity is the regulation of neutrophil elastase.⁵⁶ In addition, AAT reduces the production of pro-inflammatory cytokines, inhibits apoptosis, blocks leukocyte degranulation and migration, and modulates local and systemic inflammatory responses.⁵⁸ Inhibitory properties of this glycoprotein against serine proteases maintain the protease/antiprotease balance in the body, which is the primary mechanism protecting against uncontrolled proteolysis in tissues.⁵⁹ AAT is responsible for approximately 80-90% of antiprotease activity in blood plasma.⁶⁰ One of the essential functions of this protein is to inhibit the activity of neutrophil elastase and trypsin by forming inactive complexes with them. It also inhibits cathepsin released from neutrophils and blocks the cytotoxicity of neutrophils and their effect on IL-8 produced by epithelial cells.⁵⁷

Moreover, the presence of methionine groups in the active center of this glycoprotein indicates its possible antioxidant activity.^{61, 62}

Post-exercise Inflammation

Regular exercise generally strengthens muscles and increases their resistance to fatigue.⁶³ However, during intense exercise, tired muscles are temporarily weakened.⁶⁴ In untrained individuals, a single series of moderate to vigorous exercise can damage muscles and activate neutrophils in response to developing inflammation, ultimately causing muscle soreness.⁶⁵ It was shown that exercise produces a response similar to the acute phase in response to sepsis or mechanical tissue trauma.⁶⁶ Activation of inflammatory response is essential for muscle repair and adaptation to exercise. On the other hand, excessive or persistent inflammation is believed to contribute to further tissue damage due to nonspecific phagocytic function of inflammatory cells, such as neutrophils.⁶⁷ Therefore, reducing inflammation may have a beneficial effect on the recovery of muscle function after intense exercise.⁶⁸ Both endurance exercise and interval training induce metabolic stress in active skeletal muscles associated with a high rate of aerobic respiration and heat production.⁶⁹ After training, there is often an increase in markers of skeletal muscle damage in the blood, such as creatine kinase (CK) or lactate dehydrogenase (LDH).^{70, 71} CK is a frequently used diagnostic marker for detecting exercise-induced muscle damage.⁷² Moreover, increased serum CK activity after exercise is inversely proportional to muscular strength.^{73, 74} LDH is an enzyme that catalyzes the final step of anaerobic glycolysis, regenerating nicotinamide adenine dinucleotide reduced (NADH) to oxidized (NAD⁺) form by converting pyruvate to lactate.⁷⁵ This enzyme is involved in the metabolism of muscles, and its activity is constantly present in the blood serum as a result of energy expenditure. However, LDH activity increases significantly after intensive physical exercise.⁷⁶ Serum LDH indicates the degree of metabolic adaptation of skeletal muscles to physical training.⁷⁷ In addition, after exercise, an increase in the concentration of inflammatory markers (e.g., selected interleukins, CRP, etc.) may be observed.^{2, 78} A previous study reported increased concentrations of markers (CRP, myoglobin, CK) associated with inflammation and muscle injury in the blood sample of athletes after two ultra-endurance mountain races of different distances (54 and 111 km).⁷⁴ Regular training was shown to lower CRP levels compared to baseline.⁷⁹ This is due to

various mechanisms and processes during the adaptation of an organism to workload. A high concentration of CRP after exercise indicates a lack of adaptation to physical exertion or overtraining, primarily due to oxidative stress and inflammation.³²

Oxidative stress is also important in modulating the inflammatory response after exercise. However, the association between inflammatory mediators (e.g., cytokines) and oxidative stress is not fully explained. Increasing demand for energy after exercise causes greater oxygen consumption by mitochondria and thus increases ROS production.⁸⁰ Superoxide (O_2^-) and nitrogen monoxide (NO) are the main free radicals that trigger a chain reaction of hydrogen peroxide (H_2O_2), hydroxyl radicals (OH^\cdot), peroxynitrite ($ONOO^-$), and hypochlorous acid (HOCl). All these free radicals can cause immune reactions and cellular signaling. However, they also have negative effects, such as causing oxidative damage to lipids or proteins.⁸¹ ROS is produced by a leak of single electrons in the respiratory chain in the inner membrane of mitochondria of the contracting muscle cells.⁸² Other sources of ROS in muscle fibers during physical exercises can increase the activity of phospholipase A2 (PLA2), nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, and xanthine oxidase (XO).⁸³ NADPH oxidase is believed to increase the production of superoxides by releasing electrons, whereas XO produces superoxides by converting hypoxanthine into xanthine and uric acid.⁸¹ The main source of ROS during physical activity probably depends on the type of exercise.⁸⁴ Intensive exercise can trigger muscle injuries, which activate neutrophils and macrophages via interferon- γ (IFN- γ), IL-1, and TNF. These immune cells excessively produce ROS by oxidative burst, a main component of the neutrophil defense mechanism.⁸² NO production increases during exercise to improve vascularization and skeletal muscle function by modulating microcirculation. Consequently, superoxide anions react with NO to produce different forms of reactive nitrogen (RNS), causing further ROS generation and reducing NO bioavailability.⁸⁵ When redox homeostasis is disrupted, cells become vulnerable to ROS attack, resulting in oxidative damage to cellular components.⁸⁶ High levels of ROS lead to peroxidation of cell membranes and damage to numerous macromolecules, including DNA.⁸⁷ In addition, ROS destabilizes muscle cell structures, including the sarcolemma. Prolonged high-energy transformation to sustain repetitive contractions and increased intramuscular pressure induces mild hypoxic stress on the

muscle fibers, resulting in the accumulation of metabolites, which in turn increase the osmolality of cells.⁶⁹ Damage to the cell membrane of myocytes increases permeability to various substances.⁶⁹ When skeletal muscles are damaged, sarcolemma is destabilized, and extracellular calcium ions immediately enter the damaged cells. Subsequently, myocyte degeneration begins by activated calpain, a calcium-dependent neutral protease protein.⁸⁸ Structural changes in muscle fibers are also accompanied by increased release of some intracellular enzymes.^{89, 90} Lysosomal membranes may be damaged after exercise, resulting in lysosomal enzymes entering the cytoplasm. Subsequently, they are released from cells into the bloodstream and can contribute to inflammation.⁹¹⁻⁹³ In the acute phase, polymorphonuclear leukocytes are the most abundant cells at the injury site. However, they are replaced by monocytes during the first day of the inflammatory reaction. Based on the basic principles of inflammation, these monocytes are eventually converted into macrophages and then actively engaged in the proteolysis and phagocytosis of necrotic material by releasing lysosomal enzymes.⁹⁴ The main function of the acute phase response is to protect tissues from further injury and return to homeostasis.⁹⁵ Immune stress induces the production of pro-inflammatory cytokines that diffuse into the circulation, which in turn causes the liver to promote the secretion of APPs such as CRP, serum amyloid P (SAP), SAA, or AAT.¹⁵ Serine protease inhibitors are proteins that regulate and control crucial physiological processes, such as inflammation, coagulation, thrombosis and thrombolysis, and immune responses.⁹⁶ AAT inhibits the protease elastase produced by neutrophils during an inflammatory response. It is possible that increased AAT indirectly inhibits complement activation.⁹⁷ It was shown that AAT reduces the production of pro-inflammatory cytokines, inhibits apoptosis and inflammatory reactions.⁵⁸ Several studies reported that AAT activity is increased post-exercise.⁹⁸⁻¹⁰¹ Another study reported post-exercise increase of both IL-6 concentration and AAT activity, which may confirm that the release of pro-inflammatory cytokines stimulates the release of APPs.⁹³

Muscle activity increases ROS production and enhances the body's antioxidant defense system. Antioxidant enzymes superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPX) are the primary defense against ROS generated during exercise, as their levels increase in response to exercise.⁸² Some studies that evaluated the

effect of exercise have shown increased ROS production and antioxidant activity of enzymes.^{86, 102, 103} Ammar and colleagues reported increased SOD and GPX activity in healthy, untrained men after exercise.⁸³ Increased antioxidant activity may result from adaptation to increased ROS concentrations.¹⁰⁴ Therefore, regular exercise may improve antioxidant ability and maintain oxidant-antioxidant balance.⁸⁴ Habitual intense training is shown to reduce lipid peroxidation products and prevent oxidative damage in tissues.¹⁰⁵ Low concentrations of ROS are essential for proper regulation of cellular function and adaptation to exercise-induced stress. Well-trained athletes were shown to be more susceptible to oxidative stress.¹⁰⁶ As a result of muscle tissue damage caused by ROS, inflammatory mediators involved in initiating the inflammatory response are released at the site of damage.^{69, 107} Exercise can trigger acute phase response characterized by an increase in peripheral blood levels of cytokines and chemokines.⁸⁰

The immune system plays a significant role in the degeneration and regeneration of muscles and surrounding connective tissue after exercise-induced damage.¹⁰⁸ Cytokines play a particular role in maintaining the balance between beneficial and adverse physiological effects of the immune system response to exercise.¹⁰⁹ IL-6 has both pro- and anti-inflammatory properties,³¹ and may modulate anti-inflammatory response.³² It was shown that after intense physical exercise, the levels of TNF- α and IL-1 β increase by a factor of two, and IL-6 may increase up to 100-fold.⁷⁹ Moreover, concentrations of chemokines, IL-8, and macrophage inflammatory proteins (MIP-1 α and MIP-1 β) are elevated after intense exercise.⁷⁹ TNF- α appears to be the first systemically released cytokine, with concentrations peaking within hours after inflammatory response. Shortly after, IL-1 concentration increases followed by an increase in IL-6. Anti-inflammatory cytokines are released in response to increasing levels of pro-inflammatory cytokines to inhibit and prevent tissue damage.^{14, 110} Increased levels of IL-6, IL-10, IL-8, and inflammatory cytokines after exercise were reported.^{78, 111} On the other hand, another study reported increased plasma IL-1 β levels after acute high-intensity street dance exercise.¹⁰⁷ It was also shown that changes in TGF- β concentration can be involved in the process of human body adaptation to exercise.³⁶ Over the past decade, researchers have highlighted the biological effect of muscle-derived cytokines (i.e., myokines) in regulating cellular metabolism in skeletal muscle and adipose tissue. A small number of studies have examined the relationship between released cytokines and markers of

muscle damage to establish whether cytokines are the cause or by-product of exercise-induced muscle damage.⁶⁴

Several types of immune cells (e.g., mast cells, neutrophils, T-regulatory cells, eosinophils, CD8 T cells) were shown to infiltrate damaged skeletal muscles.¹¹² Leukocytes build up in the muscles immediately after exercise and accumulate in the extracellular space in the muscle tissue 24-48 hours after exercise.¹¹² Macrophages, the predominant leukocyte type, are observed at each time point of skeletal muscle regeneration after injury and perform specific functions throughout the process.¹¹³ Macrophages invading damaged muscle tissue produce several growth factors that stimulate and promote muscle regeneration *in vitro*. Some growth factors, such as insulin-like growth factor 1 (IGF-1) and TGF- β 1 are known to regulate the activation of satellite cells.⁸⁸ These cells are activated in response to adequate stimulus, which initiates proliferation.¹¹⁴ Subsequently, they migrate to areas where they differentiate and fuse with existing muscle fibers or form new fibers.⁸⁸ It was found that exercise can stimulate satellite cells to re-enter the cell cycle and multiply.⁶⁴ After the removal of necrotic tissue, satellite cells regenerate skeletal muscle tissue by regulating transcription factors.¹⁰⁶

Characteristics of Irisin

Skeletal muscle secretes numerous myokines, which are synthesized and secreted by myocytes in response to muscle contraction. Myokines exert an autocrine function in regulating muscle metabolism.¹¹⁵ Irisin is a novel myokine released after exercise.¹¹⁶ It is a peptide derived from proteolytic cleavage of fibronectin type III domain-containing protein 5 (FNDC5). FNDC5 is a type I transmembrane protein predominantly present in skeletal muscles.^{117, 118} Some studies suggested that irisin may be involved in the regulation of energy metabolism.^{115, 116} Rashid and others reported that this myokine improves glucose homeostasis after long-term moderate physical exercise.¹¹⁷ Several studies have suggested that irisin is directly associated with the beneficial effects of regular exercise. It is shown that exercise increases serum concentration of irisin.^{119, 120} By entering the bloodstream, irisin circulates to organs and tissues and acts as a strong inhibitor of pro-inflammatory cytokines.¹²¹ Irisin is therefore a factor that explains numerous metabolic adaptations induced by exercise. However, further studies are required to elucidate the role of irisin in regulating the production of pro-inflammatory cytokines during exercise. It was reported that irisin modulates mitochondrial

fusion, fission, and biogenesis to suppress oxidative stress.¹²² Irisin concentration at rest is positively correlated with malondialdehyde, a biomarker of oxidative stress. However, it is negatively correlated with the antioxidant protection marker, *i.e.*, the capacity to absorb oxygen radicals in response to exercise.¹²² This, in turn, clarifies the association between irisin and oxidative stress in athletes with overtraining syndrome.

Practical Implications

Skeletal muscle regeneration is a highly synchronized process involving the activation of several cellular and molecular responses.¹²³ It involves interactions between inflammatory cells, fibroblasts, and endothelial cells, as well as their secreted soluble factors.¹²⁴ In recent years, sports medicine professionals have faced new challenges as a result of sports development initiatives. Proper training programs should balance the systemic stressors experienced by athletes with personalized plans to improve performance and reduce exercise-induced stress symptoms. The use of blood biomarkers, including new myokines secreted from skeletal muscle, provides a faster and potentially more accurate method for assessing muscle damage and exercise-induced inflammation. In addition, they offer valuable insight into the relative state of recovery. However, despite their high accuracy, there is a lack of consensus about the time course and magnitude of their appearance and clearance from the blood following different types or intensities of exercise.¹²⁵ To date, there is no explicit data on the ability of organisms to regenerate and reduce inflammation. Moreover, there is no published scientific evidence to aid the selection of the most appropriate post-exercise recovery method between training sessions. Therefore, an in-depth understanding of inflammatory response due to exercise is essential.

Conclusion

The accumulation of cytokines, neutrophils, and macrophages is directly associated with muscle tissue damage. Understanding the inflammatory response and mechanism of exercise-induced myokines allows the identification of inflammatory markers involved in muscle damage and regeneration. The type of post-exercise recovery that may reduce inflammatory responses seems to be key to effective training and adaptation of muscle tissues to long-term physical exertion. Further studies are required to elucidate the exact underlying mechanisms of these beneficial effects. Based on our

current knowledge, such an approach would reduce post-exercise muscle damage, which can protect the health and physical integrity of athletes and increase their chances of achieving training goals.

Authors' Contribution

C.M-K: Research conception and supervision. M.P, C.M-K: Drafting and revising the manuscript. Both authors have approved the final version of the manuscript for publication

Conflict of Interest: None declared.

References

- 1 Salman A, Lee YH. Spiritual practices and effects of spiritual well-being and depression on elders' self-perceived health. *Appl Nurs Res.* 2019;48:68-74. doi: 10.1016/j.apnr.2019.05.018. PubMed PMID: 31266611.
- 2 Niemela M, Kangastupa P, Niemela O, Bloigu R, Juvonen T. Acute Changes in Inflammatory Biomarker Levels in Recreational Runners Participating in a Marathon or Half-Marathon. *Sports Med Open.* 2016;2:21. doi: 10.1186/s40798-016-0045-0. PubMed PMID: 27747777; PubMed Central PMCID: PMC5005625.
- 3 Bleakley C, McDonough S, Gardner E, Baxter GD, Hopkins JT, Davison GW. Cold-water immersion (cryotherapy) for preventing and treating muscle soreness after exercise. *Cochrane Database Syst Rev.* 2012;2012:CD008262. doi: 10.1002/14651858.CD008262.pub2. PubMed PMID: 22336838; PubMed Central PMCID: PMC5005625.
- 4 Vieira A, Siqueira AF, Ferreira-Junior JB, do Carmo J, Durigan JL, Blazeovich A, et al. The Effect of Water Temperature during Cold-Water Immersion on Recovery from Exercise-Induced Muscle Damage. *Int J Sports Med.* 2016;37:937-43. doi: 10.1055/s-0042-111438. PubMed PMID: 27557407.
- 5 Pfafflin A, Schleicher E. Inflammation markers in point-of-care testing (POCT). *Anal Bioanal Chem.* 2009;393:1473-80. doi: 10.1007/s00216-008-2561-3. PubMed PMID: 19104782.
- 6 Paludan SR, Pradeu T, Masters SL, Mogensen TH. Constitutive immune mechanisms: mediators of host defence and immune regulation. *Nat Rev Immunol.* 2021;21:137-50. doi: 10.1038/s41577-020-0391-5. PubMed PMID: 32782357; PubMed Central PMCID: PMC7418297.

- 7 Wang J. Neutrophils in tissue injury and repair. *Cell Tissue Res.* 2018;371:531-9. doi: 10.1007/s00441-017-2785-7. PubMed PMID: 29383445; PubMed Central PMCID: PMC5820392.
- 8 Chen L, Deng H, Cui H, Fang J, Zuo Z, Deng J, et al. Inflammatory responses and inflammation-associated diseases in organs. *Onco-target.* 2018;9:7204-18. doi: 10.18632/onco-target.23208. PubMed PMID: 29467962; PubMed Central PMCID: PMC5805548.
- 9 Medzhitov R. Origin and physiological roles of inflammation. *Nature.* 2008;454:428-35. doi: 10.1038/nature07201. PubMed PMID: 18650913.
- 10 Filep JG. Leukocytes in Inflammation, Resolution of Inflammation, Autoimmune Diseases and Cancer. *Cells.* 2021;10. doi: 10.3390/cells10071735. PubMed PMID: 34359905; PubMed Central PMCID: PMC8307052.
- 11 Lei Y, Wang K, Deng L, Chen Y, Nice EC, Huang C. Redox regulation of inflammation: old elements, a new story. *Med Res Rev.* 2015;35:306-40. doi: 10.1002/med.21330. PubMed PMID: 25171147.
- 12 Pober JS, Sessa WC. Evolving functions of endothelial cells in inflammation. *Nat Rev Immunol.* 2007;7:803-15. doi: 10.1038/nri2171. PubMed PMID: 17893694.
- 13 Germolec DR, Shipkowski KA, Frawley RP, Evans E. Markers of Inflammation. *Methods Mol Biol.* 2018;1803:57-79. doi: 10.1007/978-1-4939-8549-4_5. PubMed PMID: 29882133.
- 14 Jee H, Jin Y. Effects of prolonged endurance exercise on vascular endothelial and inflammation markers. *J Sports Sci Med.* 2012;11:719-26. PubMed PMID: 24150084; PubMed Central PMCID: PMC3763320.
- 15 Serrano I, Luque A, Aran JM. Exploring the Immunomodulatory Moonlighting Activities of Acute Phase Proteins for Tolerogenic Dendritic Cell Generation. *Front Immunol.* 2018;9:892. doi: 10.3389/fimmu.2018.00892. PubMed PMID: 29760704; PubMed Central PMCID: PMC5936965.
- 16 Krishnamoorthy S, Honn KV. Inflammation and disease progression. *Cancer Metastasis Rev.* 2006;25:481-91. doi: 10.1007/s10555-006-9016-0. PubMed PMID: 17103050.
- 17 Frangogiannis NG. Cell biological mechanisms in regulation of the post-infarction inflammatory response. *Curr Opin Physiol.* 2018;1:7-13. doi: 10.1016/j.cophys.2017.09.001. PubMed PMID: 29552674; PubMed Central PMCID: PMC5851468.
- 18 Peake JM, Della Gatta P, Suzuki K, Nieman DC. Cytokine expression and secretion by skeletal muscle cells: regulatory mechanisms and exercise effects. *Exerc Immunol Rev.* 2015;21:8-25. PubMed PMID: 25826432.
- 19 Kanda K, Sugama K, Hayashida H, Sakuma J, Kawakami Y, Miura S, et al. Eccentric exercise-induced delayed-onset muscle soreness and changes in markers of muscle damage and inflammation. *Exerc Immunol Rev.* 2013;19:72-85. PubMed PMID: 23977721.
- 20 Simbertsev A, Kozlov I. Cytokine system. In: Kamkin A, editor. *Mechanical stretch and cytokines.* Dordrecht: Springer Science+Business Media BV; 2012. doi: 10.1007/978-94-007-2004-6_1.
- 21 Remick DG. Interleukin-8. *Crit Care Med.* 2005;33:S466-7. doi: 10.1097/01.ccm.0000186783.34908.18. PubMed PMID: 16340423.
- 22 Lin J, Liu X, Zhou Y, Zhu B, Wang Y, Cui W, et al. Molecular Basis of Irisin Regulating the Effects of Exercise on Insulin Resistance. *Applied Sciences.* 2022;12:5837. doi: 10.3390/app12125837.
- 23 Sabouri M, Taghibeikzadehbadr P, Shabkhiz F, Izanloo Z, Shaghghi FA. Effect of eccentric and concentric contraction mode on myogenic regulatory factors expression in human vastus lateralis muscle. *J Muscle Res Cell Motil.* 2022;43:9-20. doi: 10.1007/s10974-021-09613-x. PubMed PMID: 35018575.
- 24 Ramezani M, Taghian F. The effect of 8 weeks aerobic training and glycogen consumption on serum apelin and insulin resistance in women with type 2 diabetes. *Iran J Diabetes Obes* 2020; 11:164–72. doi: 10.18502/ijdo.v11i3.2605.
- 25 Clark A, Huebinger RM, Carlson DL, Wolf SE, Song J. Serum Level of Musclin Is Elevated Following Severe Burn. *J Burn Care Res.* 2019;40:535-40. doi: 10.1093/jbcr/irz101. PubMed PMID: 31187123.
- 26 Dugue B, Leppanen E. Adaptation related to cytokines in man: effects of regular swimming in ice-cold water. *Clin Physiol.* 2000;20:114-21. doi: 10.1046/j.1365-2281.2000.00235.x. PubMed PMID: 10735978.
- 27 Serhan CN, Savill J. Resolution of inflammation: the beginning programs the end. *Nat Immunol.* 2005;6:1191-7. doi: 10.1038/ni1276. PubMed PMID: 16369558.
- 28 Kaur S, Bansal Y, Kumar R, Bansal G. A panoramic review of IL-6: Structure, pathophysiological roles and inhibitors. *Bioorg Med Chem.* 2020;28:115327. doi: 10.1016/j.bmc.2020.115327. PubMed PMID: 31992476.

- 29 Schett G. Physiological effects of modulating the interleukin-6 axis. *Rheumatology (Oxford)*. 2018;57:ii43-ii50. doi: 10.1093/rheumatology/kex513. PubMed PMID: 29982781.
- 30 Hirano T. IL-6 in inflammation, autoimmunity and cancer. *Int Immunol*. 2021;33:127-48. doi: 10.1093/intimm/dxaa078. PubMed PMID: 33337480; PubMed Central PMCID: PMC3083294.
- 31 Lee EC, Watson G, Casa D, Armstrong LE, Kraemer W, Vingren JL, et al. Interleukin-6 responses to water immersion therapy after acute exercise heat stress: a pilot investigation. *J Athl Train*. 2012;47:655-63. doi: 10.4085/1062-6050-47.5.09. PubMed PMID: 23182014; PubMed Central PMCID: PMC3499890.
- 32 Palacios G, Pedrero-Chamizo R, Palacios N, Maroto-Sanchez B, Aznar S, Gonzalez-Gross M, et al. Biomarkers of physical activity and exercise. *Nutr Hosp*. 2015;31:237-44. doi: 10.3305/nh.2015.31.sup3.8771. PubMed PMID: 25719791.
- 33 Cabral-Santos C, Castrillon CI, Miranda RA, Monteiro PA, Inoue DS, Campos EZ, et al. Inflammatory Cytokines and BDNF Response to High-Intensity Intermittent Exercise: Effect the Exercise Volume. *Front Physiol*. 2016;7:509. doi: 10.3389/fphys.2016.00509. PubMed PMID: 27867360; PubMed Central PMCID: PMC3083294.
- 34 Soares V, Silveira de Avelar I, Espindola Mota Venancio P, Pires-Oliveira DAA, de Almeida Silva PH, Rodrigues Borges A, et al. Acute Changes in Interleukin-6 Level During Four Days of Long-Distance Walking. *J Inflamm Res*. 2020;13:871-8. doi: 10.2147/JIR.S281113. PubMed PMID: 33204137; PubMed Central PMCID: PMC3083294.
- 35 Ma W, Xu T, Wang Y, Wu C, Wang L, Yang X, et al. The role of inflammatory factors in skeletal muscle injury. *Biotarget*. 2018;2:7. doi: 10.21037/biotarget.2018.04.01.
- 36 Kimsa M, Strzalka-Mrozik B, Kimsa M, Gola J, Kochanska-Dziurawicz A, Zebrowska A, et al. Expression pattern of the transforming growth factor beta signaling genes in human peripheral blood mononuclear cells after exercise-inflammatory aspects. *Am J Hum Biol*. 2012;24:859-62. doi: 10.1002/ajhb.22311. PubMed PMID: 22915245.
- 37 Menailo ME, Malashchenko VV, Shmarov VA, Gazatova ND, Melashchenko OB, Goncharov AG, et al. Interleukin-8 favors pro-inflammatory activity of human monocytes/macrophages. *Int Immunopharmacol*. 2018;56:217-21. doi: 10.1016/j.intimp.2018.01.036. PubMed PMID: 29414654.
- 38 Dinarello CA. Interleukin-1 in the pathogenesis and treatment of inflammatory diseases. *Blood*. 2011;117:3720-32. doi: 10.1182/blood-2010-07-273417. PubMed PMID: 21304099; PubMed Central PMCID: PMC3083294.
- 39 Cerqueira C, Manfroi B, Fillatreau S. IL-10-producing regulatory B cells and plasmacytes: Molecular mechanisms and disease relevance. *Semin Immunol*. 2019;44:101323. doi: 10.1016/j.smim.2019.101323. PubMed PMID: 31685302.
- 40 Mila-Kierzenkowska C, Wozniak A, Szpinda M, Boraczynski T, Wozniak B, Rajewski P, et al. Effects of thermal stress on the activity of selected lysosomal enzymes in blood of experienced and novice winter swimmers. *Scand J Clin Lab Invest*. 2012;72:635-41. doi: 10.3109/00365513.2012.727214. PubMed PMID: 23061673.
- 41 Simonaro CM. Lysosomes, lysosomal storage diseases, and inflammation. *Journal of Inborn Errors of Metabolism and Screening*. 2019;4. doi: 10.1177/2326409816650465.
- 42 Meyer-Schwesinger C. Lysosome function in glomerular health and disease. *Cell Tissue Res*. 2021;385:371-92. doi: 10.1007/s00441-020-03375-7. PubMed PMID: 33433692; PubMed Central PMCID: PMC3083294.
- 43 Gan X, Qiu F, Jiang B, Yuan R, Xiang Y. Convenient and highly sensitive electrochemical biosensor for monitoring acid phosphatase activity. *Sensors and Actuators B: Chemical*. 2021;332:129483.
- 44 Yan X, Xia C, Chen B, Li YF, Gao PF, Huang CZ. Enzyme Activity Triggered Blocking of Plasmon Resonance Energy Transfer for Highly Selective Detection of Acid Phosphatase. *Anal Chem*. 2020;92:2130-5. doi: 10.1021/acs.analchem.9b04685. PubMed PMID: 31850751.
- 45 Anand A, Srivastava PK. A molecular description of acid phosphatase. *Appl Biochem Biotechnol*. 2012;167:2174-97. doi: 10.1007/s12010-012-9694-8. PubMed PMID: 22684363.
- 46 de Pizzol Junior JP, Sasso-Cerri E, Cerri PS. Matrix Metalloproteinase-1 and Acid Phosphatase in the Degradation of the Lamina Propria of Eruptive Pathway of Rat Molars. *Cells*. 2018;7. doi: 10.3390/cells7110206. PubMed PMID: 30423799; PubMed Central PMCID: PMC3083294.
- 47 Yener Y, Celik I, Sur E, Ozurlu Y, Ozaydin T. Effects of long term oral acrylamide administration on alpha naphthyl acetate

- esterase and acid phosphatase activities in the peripheral blood lymphocytes of rats. *Biotech Histochem.* 2019;94:352-9. doi: 10.1080/10520295.2019.1571227. PubMed PMID: 30864862.
- 48 Kovacs Z, Jung I, Gurzu S. Arylsulfatases A and B: From normal tissues to malignant tumors. *Pathol Res Pract.* 2019;215:152516. doi: 10.1016/j.prp.2019.152516. PubMed PMID: 31262576.
- 49 Diez-Roux G, Ballabio A. Sulfatases and human disease. *Annu Rev Genomics Hum Genet.* 2005;6:355-79. doi: 10.1146/annurev.genom.6.080604.162334. PubMed PMID: 16124866.
- 50 Ghosh D. Human sulfatases: a structural perspective to catalysis. *Cell Mol Life Sci.* 2007;64:2013-22. doi: 10.1007/s00018-007-7175-y. PubMed PMID: 17558559.
- 51 Benes P, Vetvicka V, Fusek M. Cathepsin D--many functions of one aspartic protease. *Crit Rev Oncol Hematol.* 2008;68:12-28. doi: 10.1016/j.critrevonc.2008.02.008. PubMed PMID: 18396408; PubMed Central PMCID: PMCPMC2635020.
- 52 Ruiz-Blazquez P, Pistorio V, Fernandez-Fernandez M, Moles A. The multifaceted role of cathepsins in liver disease. *J Hepatol.* 2021;75:1192-202. doi: 10.1016/j.jhep.2021.06.031. PubMed PMID: 34242696.
- 53 Aghdassi AA, John DS, Sandler M, Weiss FU, Reinheckel T, Mayerle J, et al. Cathepsin D regulates cathepsin B activation and disease severity predominantly in inflammatory cells during experimental pancreatitis. *J Biol Chem.* 2018;293:1018-29. doi: 10.1074/jbc.M117.814772. PubMed PMID: 29229780; PubMed Central PMCID: PMCPMC5777244.
- 54 Fusek M, Vetvicka V. Dual role of cathepsin D: ligand and protease. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub.* 2005;149:43-50. doi: 10.5507/bp.2005.003. PubMed PMID: 16170387.
- 55 Popławska B, Janciauskiene S, Chorostowska-Wynimko J. [Genetic variants of alpha-1 antitrypsin: classification and clinical implications]. *Pneumonol Alergol Pol.* 2013;81:45-54. PubMed PMID: 23258471.
- 56 Sanders CL, Ponte A, Kueppers F. The Effects of Inflammation on Alpha 1 Antitrypsin Levels in a National Screening Cohort. *COPD.* 2018;15:10-6. doi: 10.1080/15412555.2017.1401600. PubMed PMID: 29381093.
- 57 Bergin DA, Hurley K, McElvaney NG, Reeves EP. Alpha-1 antitrypsin: a potent anti-inflammatory and potential novel therapeutic agent. *Arch Immunol Ther Exp (Warsz).* 2012;60:81-97. doi: 10.1007/s00005-012-0162-5. PubMed PMID: 22349104.
- 58 Ehlers MR. Immune-modulating effects of alpha-1 antitrypsin. *Biol Chem.* 2014;395:1187-93. doi: 10.1515/hsz-2014-0161. PubMed PMID: 24854541; PubMed Central PMCID: PMCPMC4237306.
- 59 Thompson D, Milford-Ward A, Whicher JT. The value of acute phase protein measurements in clinical practice. *Ann Clin Biochem.* 1992;29:123-31. doi: 10.1177/000456329202900201. PubMed PMID: 1378257.
- 60 Corlateanu A, Covantev S, Caraivanova I, Bodrug V, Botnaru V, Varon J, et al. Alpha-1 antitrypsin deficiency and chronic obstructive pulmonary disease: Between overlaps, phenotypes and illnesses. *Current Respiratory Medicine Reviews.* 2019;15:147-55. doi: 10.2174/1573398X15666190617143122.
- 61 Brantly M. Alpha1-antitrypsin: not just an antiprotease: extending the half-life of a natural anti-inflammatory molecule by conjugation with polyethylene glycol. *Am J Respir Cell Mol Biol.* 2002;27:652-4. doi: 10.1165/rcmb.F250. PubMed PMID: 12444023.
- 62 Levine RL, Moskovitz J, Stadtman ER. Oxidation of methionine in proteins: roles in antioxidant defense and cellular regulation. *IUBMB Life.* 2000;50:301-7. doi: 10.1080/713803735. PubMed PMID: 11327324.
- 63 Bogdanis GC. Effects of physical activity and inactivity on muscle fatigue. *Front Physiol.* 2012;3:142. doi: 10.3389/fphys.2012.00142. PubMed PMID: 22629249; PubMed Central PMCID: PMCPMC3355468.
- 64 Paulsen G, Mikkelsen UR, Raastad T, Peake JM. Leucocytes, cytokines and satellite cells: what role do they play in muscle damage and regeneration following eccentric exercise? *Exerc Immunol Rev.* 2012;18:42-97. PubMed PMID: 22876722.
- 65 Yimcharoen M, Kittikunnathum S, Suknikorn C, Nak-On W, Yeethong P, Anthony TG, et al. Effects of ascorbic acid supplementation on oxidative stress markers in healthy women following a single bout of exercise. *J Int Soc Sports Nutr.* 2019;16:2. doi: 10.1186/s12970-019-0269-8. PubMed PMID: 30665439; PubMed Central PMCID: PMCPMC6341721.
- 66 Scheffer DDL, Latini A. Exercise-induced immune system response: Anti-inflammatory status on peripheral and central organs. *Biochim Biophys Acta Mol Basis Dis.* 2020;1866:165823. doi: 10.1016/j.bbadis.2020.165823. PubMed PMID: 32360589; PubMed Central PMCID:

- PMCPMC7188661.
- 67 White G, Caterini JE. Cold water immersion mechanisms for recovery following exercise: cellular stress and inflammation require closer examination. *J Physiol*. 2017;595:631-2. doi: 10.1113/JP273659. PubMed PMID: 28145015; PubMed Central PMCID: PMCPMC5285611.
 - 68 Abaidia AE, Lamblin J, Delecroix B, Leduc C, McCall A, Nedelec M, et al. Recovery From Exercise-Induced Muscle Damage: Cold-Water Immersion Versus Whole-Body Cryotherapy. *Int J Sports Physiol Perform*. 2017;12:402-9. doi: 10.1123/ijsp.2016-0186. PubMed PMID: 27396361.
 - 69 White GE, Wells GD. Cold-water immersion and other forms of cryotherapy: physiological changes potentially affecting recovery from high-intensity exercise. *Extrem Physiol Med*. 2013;2:26. doi: 10.1186/2046-7648-2-26. PubMed PMID: 24004719; PubMed Central PMCID: PMCPMC3766664.
 - 70 Brancaccio P, Maffulli N, Limongelli FM. Creatine kinase monitoring in sport medicine. *Br Med Bull*. 2007;81-82:209-30. doi: 10.1093/bmb/ldm014. PubMed PMID: 17569697.
 - 71 Romagnoli M, Alis R, Aloe R, Salvagno GL, Basterra J, Pareja-Galeano H, et al. Influence of training and a maximal exercise test in analytical variability of muscular, hepatic, and cardiovascular biochemical variables. *Scand J Clin Lab Invest*. 2014;74:192-8. doi: 10.3109/00365513.2013.873948. PubMed PMID: 24484196.
 - 72 Reichel T, Bosslau TK, Palmowski J, Eder K, Ringseis R, Mooren FC, et al. Reliability and suitability of physiological exercise response and recovery markers. *Sci Rep*. 2020;10:11924. doi: 10.1038/s41598-020-69280-9. PubMed PMID: 32681124; PubMed Central PMCID: PMCPMC7368084.
 - 73 Fonseca LB, Brito CJ, Silva RJ, Silva-Grigoletto ME, da Silva WMJ, Franchini E. Use of Cold-Water Immersion to Reduce Muscle Damage and Delayed-Onset Muscle Soreness and Preserve Muscle Power in Jiu-Jitsu Athletes. *J Athl Train*. 2016;51:540-9. doi: 10.4085/1062-6050-51.9.01. PubMed PMID: 27575565; PubMed Central PMCID: PMCPMC5317190.
 - 74 Rubio-Arias JA, Avila-Gandia V, Lopez-Roman FJ, Soto-Mendez F, Alcaraz PE, Ramos-Campo DJ. Muscle damage and inflammation biomarkers after two ultra-endurance mountain races of different distances: 54 km vs 111 km. *Physiol Behav*. 2019;205:51-7. doi: 10.1016/j.physbeh.2018.10.002. PubMed PMID: 30291850.
 - 75 Kim JW, Dang CV. Multifaceted roles of glycolytic enzymes. *Trends Biochem Sci*. 2005;30:142-50. doi: 10.1016/j.tibs.2005.01.005. PubMed PMID: 15752986.
 - 76 Bernat-Adell MD, Collado-Boira EJ, Moles-Julio P, Panizo-Gonzalez N, Martinez-Navarro I, Hernando-Fuster B, et al. Recovery of Inflammation, Cardiac, and Muscle Damage Biomarkers After Running a Marathon. *J Strength Cond Res*. 2021;35:626-32. doi: 10.1519/JSC.0000000000003167. PubMed PMID: 31045685.
 - 77 Brancaccio P, Limongelli FM, Maffulli N. Monitoring of serum enzymes in sport. *Br J Sports Med*. 2006;40:96-7. doi: 10.1136/bjism.2005.020719. PubMed PMID: 16431993; PubMed Central PMCID: PMCPMC2492050.
 - 78 Nielsen HG, Oktedalen O, Opstad PK, Lyberg T. Plasma Cytokine Profiles in Long-Term Strenuous Exercise. *J Sports Med (Hindawi Publ Corp)*. 2016;2016:7186137. doi: 10.1155/2016/7186137. PubMed PMID: 27239554; PubMed Central PMCID: PMCPMC4864530.
 - 79 Pedersen BK, Steensberg A, Fischer C, Keller C, Ostrowski K, Schjerling P. Exercise and cytokines with particular focus on muscle-derived IL-6. *Exerc Immunol Rev*. 2001;7:18-31. PubMed PMID: 11579746.
 - 80 Accattato F, Greco M, Pullano SA, Care I, Fiorillo AS, Pujia A, et al. Effects of acute physical exercise on oxidative stress and inflammatory status in young, sedentary obese subjects. *PLoS One*. 2017;12:e0178900. doi: 10.1371/journal.pone.0178900. PubMed PMID: 28582461; PubMed Central PMCID: PMCPMC5459463.
 - 81 Kawamura T, Muraoka I. Exercise-Induced Oxidative Stress and the Effects of Antioxidant Intake from a Physiological Viewpoint. *Antioxidants (Basel)*. 2018;7. doi: 10.3390/antiox7090119. PubMed PMID: 30189660; PubMed Central PMCID: PMCPMC6162669.
 - 82 Steinbacher P, Eckl P. Impact of oxidative stress on exercising skeletal muscle. *Biomolecules*. 2015;5:356-77. doi: 10.3390/biom5020356. PubMed PMID: 25866921; PubMed Central PMCID: PMCPMC4496677.
 - 83 Ammar A, Trabelsi K, Boukhris O, Glenn JM, Bott N, Masmoudi L, et al. Effects of Aerobic-, Anaerobic- and Combined-Based Exercises on Plasma Oxidative Stress Biomarkers in Healthy Untrained Young Adults. *Int J Environ Res Public Health*. 2020;17. doi: 10.3390/ijerph17072601. PubMed PMID: 32290148; PubMed Central PMCID: PMCPMC7178085.

- 84 Sutkowy P, Wozniak A, Mila-Kierzenkowska C, Szewczyk-Golec K, Wesolowski R, Pawłowska M, et al. Physical Activity vs. Redox Balance in the Brain: Brain Health, Aging and Diseases. *Antioxidants (Basel)*. 2021;11. doi: 10.3390/antiox11010095. PubMed PMID: 35052600; PubMed Central PMCID: PMCPMC8773223.
- 85 Bosco G, Paganini M, Giacomini TA, Oppio A, Vezzoli A, Dellanoce C, et al. Oxidative Stress and Inflammation, MicroRNA, and Hemoglobin Variations after Administration of Oxygen at Different Pressures and Concentrations: A Randomized Trial. *Int J Environ Res Public Health*. 2021;18. doi: 10.3390/ijerph18189755. PubMed PMID: 34574676; PubMed Central PMCID: PMCPMC8468581.
- 86 Souissi W, Bouzid MA, Farjallah MA, Ben Mahmoud L, Boudaya M, Engel FA, et al. Effect of Different Running Exercise Modalities on Post-Exercise Oxidative Stress Markers in Trained Athletes. *Int J Environ Res Public Health*. 2020;17. doi: 10.3390/ijerph17103729. PubMed PMID: 32466187; PubMed Central PMCID: PMCPMC7277356.
- 87 Silva MA, Carvalho TR, Cruz AC, Jesus LR, Silva Neto LA, Trajano ET, et al. Effect of time-dependent cryotherapy on redox balance of quadriceps injuries. *Cryobiology*. 2016;72:1-6. doi: 10.1016/j.cryobiol.2016.01.001. PubMed PMID: 26769009.
- 88 Takagi R, Fujita N, Arakawa T, Kawada S, Ishii N, Miki A. Influence of icing on muscle regeneration after crush injury to skeletal muscles in rats. *J Appl Physiol (1985)*. 2011;110:382-8. doi: 10.1152/jappphysiol.01187.2010. PubMed PMID: 21164157.
- 89 Wozniak A, Wozniak B, Drewa G, Mila-Kierzenkowska C, Rakowski A. The effect of whole-body cryostimulation on lysosomal enzyme activity in kayakers during training. *Eur J Appl Physiol*. 2007;100:137-42. doi: 10.1007/s00421-007-0404-0. PubMed PMID: 17458576.
- 90 Kloska A, Tyłki-Szymanska A, Wegrzyn G. Lysosomal storage diseases--an overview]. *Postepy Biochem*. 2011;57:128-32. PubMed PMID: 21913413.
- 91 Sutkowy P, Woźniak A, Mila-Kierzenkowska C, Jurecka A. The activity of lysosomal enzymes in the healthy men's blood after single Finnish sauna procedure--preliminary study. *Medical and Biological Sciences*. 2012;26:33-8. doi: 10.2478/v10251-012-0052-4.
- 92 Mila-Kierzenkowska C, Woźniak A, Boraczyński T, Jurecka A, Augustyńska B, Woźniak B. The effect of whole-body cryostimulation on the activity of lysosomal enzymes in kayaker women after intense exercise. *Journal of Thermal Biology*. 2011;36:29-33. doi: 10.1016/j.jtherbio.2010.10.001.
- 93 Pawłowska M, Mila-Kierzenkowska C, Boraczynski T, Boraczynski M, Szewczyk-Golec K, Sutkowy P, et al. The Effect of Submaximal Exercise Followed by Short-Term Cold-Water Immersion on the Inflammatory State in Healthy Recreational Athletes: A Cross-Over Study. *J Clin Med*. 2021;10. doi: 10.3390/jcm10184239. PubMed PMID: 34575347; PubMed Central PMCID: PMCPMC8468461.
- 94 Jarvinen TA, Jarvinen TL, Kaariainen M, Kalimo H, Jarvinen M. Muscle injuries: biology and treatment. *Am J Sports Med*. 2005;33:745-64. doi: 10.1177/0363546505274714. PubMed PMID: 15851777.
- 95 Wigmore SJ, Fearon KC, Ross JA, McNally SJ, Welch WJ, Garden OJ. Febrile-range temperature but not heat shock augments the acute phase response to interleukin-6 in human hepatoma cells. *Am J Physiol Gastrointest Liver Physiol*. 2006;290:G903-11. doi: 10.1152/ajpgi.00089.2005. PubMed PMID: 16339299.
- 96 Kellici TF, Pilka ES, Bodkin MJ. Small-molecule modulators of serine protease inhibitor proteins (serpins). *Drug Discov Today*. 2021;26:442-54. doi: 10.1016/j.drudis.2020.11.012. PubMed PMID: 33259801.
- 97 Vassalle C, Masotti S, Lubrano V, Basta G, Prontera C, Di Cecco P, et al. Traditional and new candidate cardiac biomarkers assessed before, early, and late after half marathon in trained subjects. *Eur J Appl Physiol*. 2018;118:411-7. doi: 10.1007/s00421-017-3783-x. PubMed PMID: 29256048.
- 98 Markovitch D, Tyrrell RM, Thompson D. The effect of prior exercise on ex vivo induction of heme oxygenase-1 in human lymphocytes. *Free Radic Res*. 2007;41:1125-34. doi: 10.1080/10715760701589230. PubMed PMID: 17886034.
- 99 Semple SJ, Smith LL, McKune AJ, Hoyos J, Mokgethwa B, San Juan AF, et al. Serum concentrations of C reactive protein, alpha1 antitrypsin, and complement (C3, C4, C1 esterase inhibitor) before and during the Vuelta a Espana. *Br J Sports Med*. 2006;40:124-7. doi: 10.1136/bjism.2005.019489. PubMed PMID: 16431998; PubMed Central PMCID: PMCPMC2492037.
- 100 Schild M, Eichner G, Beiter T, Zugel M, Krumholz-Wagner I, Hudemann J, et al.

- Effects of Acute Endurance Exercise on Plasma Protein Profiles of Endurance-Trained and Untrained Individuals over Time. *Mediators Inflamm.* 2016;2016:4851935. doi: 10.1155/2016/4851935. PubMed PMID: 27239103; PubMed Central PMCID: PMCPMC4867072.
- 101 Powers SK, Schrager M. Redox signaling regulates skeletal muscle remodeling in response to exercise and prolonged inactivity. *Redox Biol.* 2022;54:102374. doi: 10.1016/j.redox.2022.102374. PubMed PMID: 35738088; PubMed Central PMCID: PMCPMC9233275.
- 102 Inal M, Akyuz F, Turgut A, Getsfrid WM. Effect of aerobic and anaerobic metabolism on free radical generation swimmers. *Med Sci Sports Exerc.* 2001;33:564-7. doi: 10.1097/00005768-200104000-00009. PubMed PMID: 11283431.
- 103 Bouzid MA, Filaire E, Matran R, Robin S, Fabre C. Lifelong Voluntary Exercise Modulates Age-Related Changes in Oxidative Stress. *Int J Sports Med.* 2018;39:21-8. doi: 10.1055/s-0043-119882. PubMed PMID: 29169189.
- 104 Lu Y, Wiltshire HD, Baker JS, Wang Q. Effects of High Intensity Exercise on Oxidative Stress and Antioxidant Status in Untrained Humans: A Systematic Review. *Biology (Basel).* 2021;10. doi: 10.3390/biology10121272. PubMed PMID: 34943187; PubMed Central PMCID: PMCPMC8698973.
- 105 Castrogiovanni P, Imbesi R. Oxidative stress and skeletal muscle in exercise. *Ital J Anat Embryol.* 2012;117:107-17. PubMed PMID: 23420998.
- 106 Slattery K, Bentley D, Coutts AJ. The role of oxidative, inflammatory and neuroendocrinological systems during exercise stress in athletes: implications of antioxidant supplementation on physiological adaptation during intensified physical training. *Sports Med.* 2015;45:453-71. doi: 10.1007/s40279-014-0282-7. PubMed PMID: 25398224.
- 107 Borges L, Dermargos A, Gray S, Barros Silva MB, Santos V, Pithon-Curi TC, et al. Neutrophil Migration and Adhesion Molecule Expression after Acute High-Intensity Street Dance Exercise. *J Immunol Res.* 2018;2018:1684013. doi: 10.1155/2018/1684013. PubMed PMID: 30069484; PubMed Central PMCID: PMCPMC6057282.
- 108 Peake J, Nosaka K, Suzuki K. Characterization of inflammatory responses to eccentric exercise in humans. *Exerc Immunol Rev.* 2005;11:64-85. PubMed PMID: 16385845.
- 109 Andersson H, Bohn SK, Raastad T, Paulsen G, Blomhoff R, Kadi F. Differences in the inflammatory plasma cytokine response following two elite female soccer games separated by a 72-h recovery. *Scand J Med Sci Sports.* 2010;20:740-7. doi: 10.1111/j.1600-0838.2009.00989.x. PubMed PMID: 19765242.
- 110 Suzuki K. Cytokine response to exercise and its modulation. *Antioxidants.* 2018;7:17. doi: 10.3390/antiox7010017. PubMed Central PMCID: PMC5789327.
- 111 Kouvelioti R, Kurgan N, Falk B, Ward WE, Josse AR, Klentrou P. Cytokine and Sclerostin Response to High-Intensity Interval Running versus Cycling. *Med Sci Sports Exerc.* 2019;51:2458-64. doi: 10.1249/MSS.0000000000002076. PubMed PMID: 31246713.
- 112 Peake JM, Neubauer O, Della Gatta PA, Nosaka K. Muscle damage and inflammation during recovery from exercise. *J Appl Physiol (1985).* 2017;122:559-70. doi: 10.1152/jappphysiol.00971.2016. PubMed PMID: 28035017.
- 113 Chazaud B. Inflammation during skeletal muscle regeneration and tissue remodeling: application to exercise-induced muscle damage management. *Immunol Cell Biol.* 2016;94:140-5. doi: 10.1038/icb.2015.97. PubMed PMID: 26526620.
- 114 Crameri RM, Langberg H, Magnusson P, Jensen CH, Schroder HD, Olesen JL, et al. Changes in satellite cells in human skeletal muscle after a single bout of high intensity exercise. *J Physiol.* 2004;558:333-40. doi: 10.1113/jphysiol.2004.061846. PubMed PMID: 15121802; PubMed Central PMCID: PMCPMC1664917.
- 115 Gomarasca M, Banfi G, Lombardi G. Myokines: The endocrine coupling of skeletal muscle and bone. *Adv Clin Chem.* 2020;94:155-218. doi: 10.1016/bs.acc.2019.07.010. PubMed PMID: 31952571.
- 116 Laurens C, Bergouignan A, Moro C. Exercise-Released Myokines in the Control of Energy Metabolism. *Front Physiol.* 2020;11:91. doi: 10.3389/fphys.2020.00091. PubMed PMID: 32116795; PubMed Central PMCID: PMCPMC7031345.
- 117 Rashid FA, Abbas HJ, Naser NA, Addai Ali H. Effect of Long-Term Moderate Physical Exercise on Irisin between Normal Weight and Obese Men. *Scientific-WorldJournal.* 2020;2020:1897027. doi: 10.1155/2020/1897027. PubMed PMID: 32952453; PubMed Central PMCID: PMCPMC7481929.

- 118 Sousa RAL, Improtta-Caria AC, Souza BSF. Exercise-Linked Irisin: Consequences on Mental and Cardiovascular Health in Type 2 Diabetes. *Int J Mol Sci*. 2021;22. doi: 10.3390/ijms22042199. PubMed PMID: 33672171; PubMed Central PMCID: PMC7926886.
- 119 Fatouros IG. Is irisin the new player in exercise-induced adaptations or not? A 2017 update. *Clin Chem Lab Med*. 2018;56:525-48. doi: 10.1515/cclm-2017-0674. PubMed PMID: 29127759.
- 120 Fox J, Rioux BV, Goulet EDB, Johanssen NM, Swift DL, Bouchard DR, et al. Effect of an acute exercise bout on immediate post-exercise irisin concentration in adults: A meta-analysis. *Scand J Med Sci Sports*. 2018;28:16-28. doi: 10.1111/sms.12904. PubMed PMID: 28453881.
- 121 Zhao J, Qiao L, Dong J, Wu R. Antioxidant Effects of Irisin in Liver Diseases: Mechanistic Insights. *Oxid Med Cell Longev*. 2022;2022:3563518. doi: 10.1155/2022/3563518. PubMed PMID: 35035659; PubMed Central PMCID: PMC8759828.
- 122 Joro R, Korkmaz A, Lakka TA, Uusitalo ALT, Atalay M. Plasma irisin and its associations with oxidative stress in athletes suffering from overtraining syndrome. *Physiol Int*. 2020;107:513-26. doi: 10.1556/2060.2020.00037. PubMed PMID: 33393937.
- 123 Karalaki M, Fili S, Philippou A, Koutsilieris M. Muscle regeneration: cellular and molecular events. *In Vivo*. 2009;23:779-96. PubMed PMID: 19779115.
- 124 Peake JM, Roberts LA, Figueiredo VC, Egner I, Krog S, Aas SN, et al. The effects of cold water immersion and active recovery on inflammation and cell stress responses in human skeletal muscle after resistance exercise. *J Physiol*. 2017;595:695-711. doi: 10.1113/JP272881. PubMed PMID: 27704555; PubMed Central PMCID: PMC5285720.
- 125 Bessa AL, Oliveira VN, Agostini GG, Oliveira RJ, Oliveira AC, White GE, et al. Exercise Intensity and Recovery: Biomarkers of Injury, Inflammation, and Oxidative Stress. *J Strength Cond Res*. 2016;30:311-9. doi: 10.1519/JSC.0b013e31828f1ee9. PubMed PMID: 23604000.