The Protective Effect of Taurine and Curcumin on Autophagy-Related Genes in the Oocytes of the Mouse Treated with Acrylamide

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What's Known

 Acrylamide exerts its effects through the induction of oxidative stress, which results in autophagy. High levels of oxidative stress can cause DNA damage and lipid peroxidation, which critically affects the etiology of infertility.

What's New

 Acrylamide negatively impacts oocyte viability and causes autophagy-related genes to be expressed at higher levels. It was shown that taurine was more effective than curcumin at reducing the harmful effects of acrylamide. As a result, curcumin should be used with greater caution during the first trimester of pregnancy.

Abstract

Background: Autophagy is also essential for both male and female infertility since it controls the development of germ cells and reproductive organs. This study aimed to investigate the effects of taurine and curcumin on the expression of genes related to autophagy in acrylamide-treated mice.

Methods: In 2022, this experimental study was conducted at the Shiraz University of Medical Sciences, Stem Cells Technology Research Centre. Forty-eight mice were randomly assigned to eight groups (control, curcumin 200 mg/Kg, taurine 150 mg/ Kg, acrylamide 50 mg/Kg, acrylamide+curcumin 100 mg/Kg, acrylamide+curcumin 200, acrylamide+taurine 75 mg/Kg, acrylamide+taurine 150 mg/Kg). Finally, oocyte characteristics and gene expression were determined in each group using oneway analysis of variance (ANOVA) by SPSS 25 and GraphPad 9, respectively. P<0.05 was conducted statistically significant. **Results:** A significant decrease was observed in several oocytes in the acrylamide group compared to the control group (P<0.001). The expression levels of light chain 3 (LC3), autophagy-related gene (ATG)12, ATG5, and Beclin1 significantly increased in the acrylamide compared to the control group. A significant increase in the number of oocytes was observed in the taurine group compared to the control. The expression levels of LC3, ATG12, ATG5, and Beclin1 significantly decreased in the acrylamide+taurine (150) mg/Kg) compared to the acrylamide group.

Conclusion: The acrylamide negatively impacts oocyte viability and causes the higher expression of autophagy-related genes. Taurine may encourage the fusion of autophagosomes with lysosomes by removing autophagic obstruction, potentially accelerating autophagy and protecting against oxidative stress. Taurine is more effective than curcumin at reducing the harmful effects of acrylamide. As a result, taurine can be proposed as a potential treatment drug for acrylamide-induced infertility.

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Keywords • Acrylamide • Oocyte • Mitochondria • Autophagy • Taurin • Curcumin

Introduction

Acrylamide (ACR) is a widely used chemical in food packaging, textile production, adhesives, contact lenses, wastewater treatment, cosmetics (skin creams), drinking water, and tobacco and plastic industries. The primary dietary sources

of acrylamide include potato products, grain and cereal products, dried products, roasted nuts, and coffee.1 Acrylamide, by disrupting the sex hormone system, causes ovarian tissue damage and has detrimental effects on fertility. Acrylamide exerts its effects through the induction of oxidative stress, which results in autophagy.2 Treatment of rats with high doses of ACR increased the expression of autophagyrelated genes such as autophagy-related 12 (ATG12), ATG5, and light chain 3 (LC-3).3,4 As the main cellular mechanism in many organs' reactions to acrylamide exposure, autophagy is essential for the regulation of early reproductive processes in the reproductive system, including gamete maturation and fertilization.5 The process of autophagy generates a catabolic pathway that uses the lysosome to break down proteins and cellular organelles. The autophagy of intracellular proteins and organelles saves energy, allowing the body to adapt to changing conditions.6

The development of autophagosomes and signaling pathways depends on autophagy-related genes. *Beclin-1* is a known gene in the autophagy pathway that promotes the autophagosome's nucleation stage and links other relevant proteins to autophagy. Along with *Atg16L* and *Atg12*, *Atg5* is involved in forming membrane curvature and autophagosome maturation.⁷

High levels of oxidative stress can cause DNA damage and lipid peroxidation, which critically affects the etiology of infertility.8 Studies demonstrated that using antioxidants could be a suitable solution for mitigating the adverse effects of reactive oxygen species (ROS).9, 10

Taurine (Tau), a derivative of the amino acid cysteine, is the most prevalent non-protein amino acid in the human body. Numerous investigations have demonstrated that Tau's antioxidative, anti-cell death and anti-inflammatory properties contribute to its ability to prevent xenobiotic-induced reproductive damage. The precise targets of Tau throughout the intracellular routes are linked to mitochondrial functionality. Furthermore, exogenous Tau supplementation is beneficial for several disorders.^{11, 12}

Tau has been shown to enhance energy metabolism and mitochondrial function significantly.¹³ Additionally, Tau may significantly reduce ROS produced by mitochondria.¹⁴ It is also clear that mitochondria emit mediators of cell death and repress mitochondria-mediated cell death.¹⁵

Curcumin, another antioxidant molecule, is the main active ingredient of turmeric. It has antioxidant and anti-inflammatory properties; however, conflicting studies persist on its effects on ovarian tissue.16, 17 Curcumin has been thoroughly investigated and is thought to be an anti-cancer agent for several types of cancer, including skin, stomach, liver, lung, and colon cancers. It is also an active component with anti-inflammatory, antioxidation, antibacterial, and antifungal properties.18 antiviral, Furthermore, studies in biology and medicine have demonstrated that curcumin is a safe and efficient treatment for cancer, particularly stomach cancer. However, due to curcumin's instability in alkaline conditions and limited solubility in water and acidic pH, it has not been widely employed in oral medications.^{18, 19} Taurine and curcumin can both be considered therapeutic candidates for treating autophagyrelated female infertility. As there have been few studies on the effects of ACR on the expression of genes involved in the autophagy process in mouse oocytes, the purpose of this study was to examine the effects of taurine and curcumin on the expression of autophagy-related genes in the mouse oocytes under acrylamide treatment.

Materials and Methods

This experimental study was conducted in 2022 at Shiraz University of Medical Sciences (Shiraz, Iran) Stem Cells Technology Research Center. The procedures of this experiment were approved by the Research Ethics Committees of the Islamic Azad University Shiraz Branch (IR. IAU.SHIRAZ.REC.1402.041).

Animals

The experimental animals used in this study were 48 adult female mice from the Naval Medical Research Institute, with an average weight of 30-35 g, aged 6-8-week-old. They were selected randomly and kept under standard conditions for 2 weeks before treatment to ensure adaptation. The animal room was provided with a 12-hour light-dark cycle, and a controlled temperature (20±2 °C). Ad libitum access to water and standard rat chow (Behparvar®, Tehran, Iran) was provided.

Experimental Design

The mice were grouped equally into 8 (n=6), and treated as follows:

Group 1 (Control) received physiological serum, and Group 2, 3, and 4 were treated with 200 mg/Kg curcumin (Merck-German, s7359554), 150 mg/Kg taurine (Sigma, T0625-100G, UK), and 50 mg/Kg acrylamide (Merck-German, 79-06-1), respectively. Group 5 and 6 received acrylamide+100 mg/Kg curcumin and

acrylamide+200 mg/Kg, respectively. Groups 7 and 8 received acrylamide+75mg/Kg taurine and acrylamide+150 mg/Kg taurine, respectively. The materials, including physiological serum, acrylamide, taurine, and curcumin, were administered daily for 35 days via oral gavage using a gastric needle. It is worth noting that taurine and curcumin were administered one hour before acrylamide administration.

Oocyte Collection

Female mice were super-ovulated with an intraperitoneal (i.p.) injection of 10 IU PMSG (Gonaser, HIPRA, Spain), followed by an i.p. injection of 10 IU hCG (Organon, Oss, Netherlands) on the last administration day. Between 10-12 hours post-HCG injection, the mice were euthanized by cervical dislocation, and then the oocytes were retrieved from the ampulla region of the oviduct. After briefly incubating the oocytes in 1 mg/mL hyaluronidase (4272, Sigma, UK), the cumulus cells were removed. Then, the cumulus-free oocytes were washed in preincubated fresh M2 (5910, Sigma, UK) media three times.21 Following the oocyte collection, the effects of acrylamide, taurine, and curcumin on oocytes in different groups were assessed by inverted microscopy (Nikon, Japan), and autophagy gene expression was evaluated by real-time polymerase chain reaction (real-time PCR).

Evaluation of the Level of Autophagy Gene Expression

Initially, total RNA was extracted from the oocytes (n=20 in each group) using TRIzol reagent (15-596-026 Invitrogen, USA).²² Next, the total concentration of RNA was determined using a NanoDrop (NanoDrop[™], Thermo Fisher Scientific, Wilmington, DE, USA). Then, the first strand of cDNA was synthesized using a Takara kit (Prime ScriptTM RT reagent Kit, Takara: RR037B, AUS) according to the manufacturer's instructions. All samples were

prepared for quantitative real-time PCR (gRT-PCR) conditions. The expression of Atg-5, Atg12, Lc3, and Beclin-1 genes were evaluated by Sybr Green (SG) Premix EX Taq™ II (RR650A) using the Applied Biosystems StepOnePlus™ Real-Time PCR System (Applied Biosystems, USA).²³ The primers were designed using the gene bank Primer-BLAST online program (table 1). The standard curve method was used to estimate the efficiency of a real-time PCR reaction amplification (data is not shown here) and then examine the expression of each gene in the samples. β-actin, as an internal control, was used to normalize the amplification signals of samples. All experiments were performed in triplicate, and relative expression was calculated using the 2-DACt method, which compared it with the corresponding control group.

Statistical Analysis

Version 23 of the SPSS software (Armonk, NY: IBM Corp, USA) was used for statistical analysis. First, all data were normalized, then One-way analysis of variance (ANOVA) Tukey's multiple comparison test was used to examine the data in the desired groups by GraphPad Prism software version 9 (GraphPad Software Inc. La Jolla, California, USA). Data were considered significant at a level of P<0.05.

Results

Oocyte Assessment

According to figure 1, the mean number of oocytes, oocyte diameter, zona pellucida, and perivitelline space (PVS) in the group that received acrylamide showed a significant decrease compared to the control group (P<0.001). A significant increase in the mean number of oocytes was observed in the taurine group compared to the control group (P<0.001). Additionally, a significant increase in the mean number of oocytes was observed

Table 1: Primer sequences for the forward and reverse genes involved in the autophagy process			
Gene	Sequence	Size (bp)	Annealing Temperature
M-Atg5	Forward: AACTGAAAGAGAAGCAGAACCA	105	60
M-Atg5	Reverse:TGTCTCATAACCTTCTGAAAGTGC		
M-beclin 1	Forward:AATCTAAGGAGTTGCCGTTATAC	187	60
M-beclin 1	Reverse:CCAGTGTCTTCAATCTTGCC		
M-B-actin	Forward:AGTGTGACGTTGACATCCGT	120	60
M-B-actin	Reverse:TGCTAGGAGCCAGAGCAGTA		
Atg8	Forward:GACCATCCAAGGACTCATTGAC	131	60
Atg8	Reverse:GGGCAAAGGACTGATTCACATA		
LC3	Forward:CCACCAAGATCCCAGTGATTATAG	125	60
LC3	Reverse:CAAGCGCCGTCTGATTATCT		

M: Mouse; bp: Base pair; Atg: Autophagy-related gene; LC3: Light chain 3

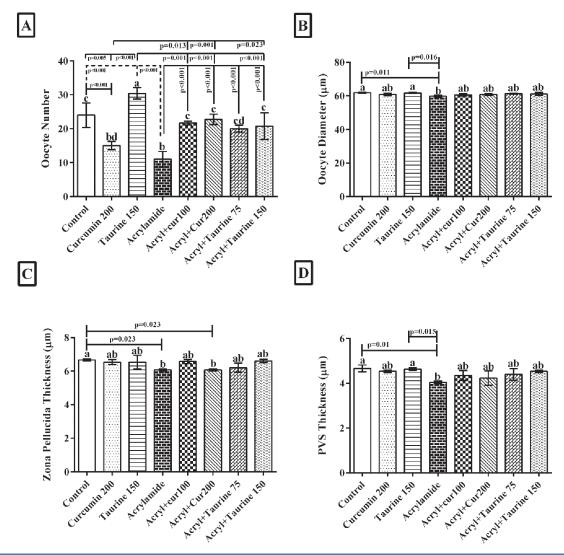


Figure 1: The number, diameter, thickness of the zona pellucida, and PVS of the oocyte in the studied groups. Data are presented as mean±SD, n=12. a.b.cThere were no significant differences between the columns containing at least one similar letter. However, different letters reveal a significant difference (P<0.05).

in the group receiving acrylamide+curcumin 100 and 200 mg/Kg and the group that received acrylamide+taurine 75 and mg/Kg compared to the group that received acrylamide alone (P<0.001). There was no significant difference in the mean number of oocytes between the groups that received acrylamide+taurine 150 mg/Kg and those receiving acrylamide+curcumin 100 and 200 mg/Kg. The oocyte diameter was significantly increased in taurine 150 mg/Kg compared to acrylamide (P<0.001). Furthermore, in the acrylamide groups treated with curcumin 100 and 200 mg/Kg and taurine 75 and 150 mg/Kg, no significant difference was observed in the mean of oocyte diameter.

The mean thickness of the zona pellucida in the groups that received acrylamide and acrylamide+curcumin 200 mg/Kg showed a significant decrease compared to the control group (P=0.023).

The PVS thickness was significantly decreased in the acrylamide groups in comparison to the control group (P<0.001).

Autophagy Gene Expression Analysis

According to figure 2A, a significant upregulation in the expression levels of the *ATG5* gene was observed in the oocytes of the acrylamidetreated group compared to other groups. The acrylamide+curcumin 100 and 200 mg/Kg groups also showed a significant increase in the expression level of *ATG5* genes compared to control groups (P=0.001 and P=0.008). The groups receiving acrylamide+curcumin 100 and 200 mg/Kg showed a significant increase in *ATG5* gene expression compared with taurine group (P=0.001 and P=0.007, respectively). There was no significant difference in the expression level of *ATG5* gene in oocytes of the control groups and taurine and curcumin groups (P>0.999 and P=0.149, respectively).

The expression level of the ATG12 gene also

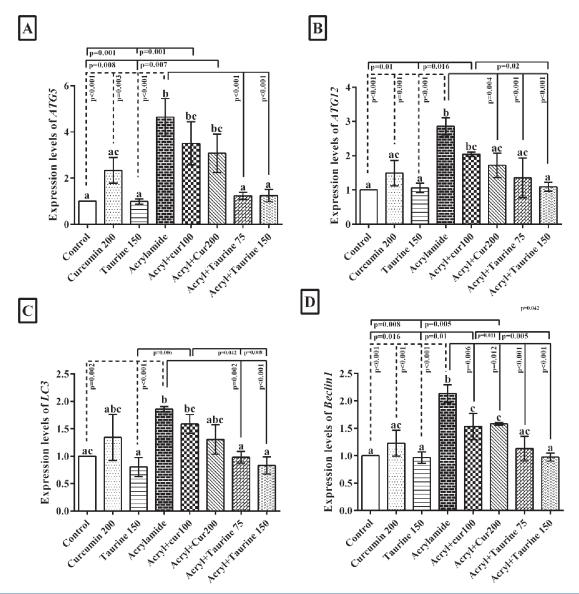


Figure 2: The related mRNA expression of ATG5, ATG12, LC3, and Beclin1 genes in oocytes in different groups including (control, curcumin 200 mg/Kg, taurine 150 mg/Kg, acrylamide 50 mg/Kg, acrylamide 50 mg/Kg+curcumin 100 mg/Kg, acrylamide 50 mg/Kg+curcumin 200, acrylamide 50 mg/Kg+taurine 75 mg/Kg, acrylamide 50 mg/Kg+taurine 150). Data are presented as mean±SD, n=12. a.b.cThere were no significant differences between the columns containing at least one similar letter. However, different letters reveal a significant difference (P<0.05).

showed a significant increase in the acrylamidetreated group compared to other groups (P=0.001), but the acrylamide+curcumin 100 mg/Kg group was not significant with acrylamide group (P=0.057). significant increase in ATG12 gene expression was observed in oocytes from the acrylamide+curcumin 100 mg/Kg groups compared with the acrylamide+taurine 150 mg/ Kg group (P=0.021). However, this difference was not significant in the other treatment groups. The acrylamide+curcumin 100 mg/Kg group (P=0.01) showed a substantial increase in ATG12 gene expression compared to the control group. No significant differences were observed in ATG12 gene expression levels in oocytes from the control group and the taurine and curcumin

groups (P>0.999 and P=0.484) (figure 2B).

A significant increase in LC3 gene expression in oocytes was observed in the treatment group receiving acrylamide+curcumin at 100 mg/Kg compared to the group receiving acrylamide+taurine at 150 mg/ Kg and 75 mg/Kg at the level of (P=0.008 and P=0.042, respectively). The group receiving acrylamide+curcumin at 100 mg/ Kg (P=0.051) showed a significant increase in LC3 gene expression compared to the control group. Additionally, the group receiving acrylamide+curcumin at 100 mg/Kg (P=0.006) showed a significant increase in LC3 gene expression compared to the healthy group receiving taurine. No significant difference in

LC3 gene expression in oocytes was observed between the control group and the healthy group receiving taurine and curcumin (P=0.933 and P=0.510, respectively).

The expression level of the *Beclin1* gene showed a significant increase in the oocytes of the acrylamide-treated group compared to control, curcumin, taurine, acrylamide+taurine 75, 150 mg/Kg groups (P=0.001) (figure 2C). The acrylamide+curcumin 100 and 200 mg/Kg groups also showed a significant increase compared to the acrylamide group (P=0.006 and P=0.012, respectively).

A significant decrease in Becline 1 gene expression was shown in acrylamide+taurine groups compared mg/Kg with acrylamide+curcumin 100 mg/Kg (P=0.011) group and the acrylamide+curcumin 200 mg/ Kg (P=0.005) group. The acrylamide+curcumin mg/Kg (P=0.016) group and acrylamide+curcumin 200 mg/Kg (P=0.008) group showed a significant increase in Becline 1 gene expression compared with the control group. Additionally, the acrylamide+curcumin (100 mg/ Kg) group (P=0.01) and the acrylamide+curcumin 200 mg/Kg (P=0.005) group showed a significant increase in Becline 1 gene expression compared with the taurine group. There was no significant difference in the expression level of the Becline 1 gene in oocytes between the control and the taurine and curcumin groups (P>0.999 and P=0.662) (figure 2D).

The results indicated a significant reduction in the expression levels of LC3, ATG12, ATG5, and Beclin1 genes in the oocytes of the group that received acrylamide+taurine 75 and 150 mg/Kg compared to the group that received acrylamide alone (P<0.001). The results also showed a significant decrease in the expression levels of Beclin1 (P=0.011 and P=0.05) and ATG5 (P=0.004 and P=0.021) genes in the group that received acrylamide+taurine 150 mg/Kg compared to the group receiving acrylamide+curcumin 100 and 200 mg/Kg. Additionally, a significant reduction was observed in the expression levels of LC3 and ATG12 genes in the group that received acrylamide+taurine 150 mg/Kg compared to the group that received acrylamide+curcumin 100 mg/Kg (P=0.008 and P=0.20, respectively).

Discussion

The results of this investigation demonstrated that, in comparison to the control group, acrylamide significantly decreased the number of oocytes, oocyte diameter, zona pellucida thickness, PVS, and significantly increased the

expression of the genes *LC3*, *ATG12*, *ATG5*, and *Beclin1*. When taurine was administered instead of curcumin, the mice who received acrylamide fared better. The taurine group had a significantly larger average number of oocytes than the control group. A significant increase in the average number of oocytes was observed in the taurine group compared to the curcumin group. Numerous intracellular processes have been identified in the biological system to adapt to different types of cell stress including the production of some genes and proteins linked to antioxidant enzymes at altered levels, elevated heat shock protein levels, metallothionein synthesis, apoptosis, and autophagy.^{24,25}

Song and colleagues discovered acrylamide reduces the amounts of glycolytic intermediates, which in turn reduces glycolysis/ gluconeogenesis. raises ROS increases apoptosis, and inhibits autophagy. They identified the fundamental impacts of ACR on autophagy in their investigation. ACR suppresses autophagy by obstructing autophagic flow, as suggested by the significant increase of LC3-II and p62 levels. According to the results of metabolomics, exposure to ACR decreased the pace at which glycolysis and the TCA cycle were metabolized, while, on the other hand, increasing the metabolism of fatty acids and amino acids. In addition, ACR promoted inflammation, cell death, and ROS generation.²⁶

Natural products such as curcumin and taurine can be used in an autophagy-dependent manner to increase the rate of embryo implantation. Only a small number of research have, however, examined autophagy as a key mechanism behind the enhancement of embryo implantation and, consequently, the use of natural products in the treatment of female infertility.²⁷ The novelty of this study is using a combination of these natural products and evaluation of autophagy-related genes in oocytes.

A study showed that adding taurine to the culture medium enhances the rate of fertilization and blastocyst development when compared to the control group.28 These outcomes are in agreement with our investigation. According to this study, a significant increase in the mean number of oocytes was observed in the taurine group compared to the control group. Together, curcumin's numerous biological activities, low toxicity, and reduced side effects over synthetic drugs make it a potentially helpful treatment for female reproductive disorders, contraception, chemotherapy chemo-preventive and or treatment, and safeguards against lowering semen quality parameters.29 In normal cells, curcumin mainly induces autophagy through

different mechanisms to promote cell survival. Meanwhile, in cancer cells, curcumin causes excessive autophagy stimulation,30 leading to cell death.31 In the present study, curcumin, especially at high doses, caused a decrease in the number of oocytes, and to some extent, the expression of autophagy pathway genes. Therefore, it can be stated that curcumin in high doses leads the cell toward the apoptotic pathway, which requires further studies in this regard. Curcumin therapy at doses of 100 and 200 mg/Kg effectively restores antioxidant gene expression, with a substantial improvement at the 200 mg/Kg dose over the 100 mg/Kg dose. Several studies have demonstrated the efficiency of curcumin in alleviating oxidative stressrelated problems, including parameters such as total antioxidant capacity, malondialdehyde, and superoxide dismutase under physiological settings. 10, 32

Gao and colleagues found that taurine can prevent autophagy by activating the mammalian rapamycin (mTOR)-peroxisome target of proliferator-activated receptor-v signaling pathway. When calcium oxalate crystals damage the epithelial cells of renal tubules, taurine can activate the Akt/mTOR (protein kinase B known as Akt) signaling pathway to block ROSdependent autophagy.33 Taurine also reduces the production of LC3-II protein and green fluorescent protein -LC3 in porcine kidney 15 cells, which decreases the autophagy effects caused by Hercotoxin A.34 In the present study, significant reductions in LC3 expression were observed in the group treated with 75 mg/Kg and 150 mg/Kg taurine, indicating a decrease in the number of autophagosomes. This finding is consistent with previous studies in that taurine can inhibit autophagy. 35, 36 Thus, compared to the acrylamide group, the simultaneous treatment of taurine and acrylamide resulted in a significant reduction in the number of autophagosomes and a considerable decrease in the expression of autophagy-related genes Atg5, Atg12, LC3, and Beclin-1, suggesting that it can reduce the autophagic blockade caused by acrylamide.

According to a 2020 study by Aldawood and others., there was a considerable increase in the mRNA levels of the *ATG5* and *ATG12* genes in ovarian tissue following high-dose acrylamide injection.³ These findings align with Tan and colleagues' investigation.³⁷ However, despite autophagosomes in acrylamide-treated animals as shown by electron microscopy sections, acrylamide treatment had no effect on *LC3*. The function of the autophagic mechanism in defending the oocytes was also covered in this investigation.³⁸ Despite this, our results showed

that LC3 levels increased when acrylamide was administered. This is significant since LC3 is an autophagy marker protein mainly involved in the maturation, formation, and destruction of autophagosomes.²⁸ Taken together, these results demonstrate that autophagy affects the oocyte by inducing apoptosis in the corpus luteum or follicular atresia analysis process. It has been established that gonadotropin is required for both processes to occur in the ovaries of mice and that autophagic activation is facilitated by the estrogen receptors (ER1 and ER2).28 Based on these findings, we propose that the autophagy process works with granulosa cell death and apoptosis caused by acrylamide. According to Yang and colleagues, these substances can decrease anti-autophagic proteins including Akt and mTOR4 in ovarian cells while simultaneously inducing autophagy repair and activating the adenosine monophosphate-activated protein kinase pathway.39 On the other hand, autophagy can be induced by high doses of acrylamide treatment, as evidenced by an increase in the mRNA levels of autophagy-related genes; however, autophagy inhibition may eventually result in cell death.40 This autophagy blockage can happen by inhibiting either autophagosomelysosome fusion or lysosomal degradation, which calls for more research in this field.

Our study was limited in that protein expression by western blot was not assessed due to insufficient budget. Furthermore, the development of fertilized oocytes to the blastocyst stage should be evaluated in conjunction with the expression of *ATG* genes at each stage in the future. However, further studies are needed in this area.

Conclusion

The results indicated that acrylamide negatively impacts oocyte viability and that autophagy-related genes are expressed at higher levels. Taurine may stimulate autophagosomes to fuse with lysosomes by removing autophagic obstruction, thereby potentially accelerating the autophagy process and protecting against oxidative stress. Taurine is more effective than curcumin at reducing the harmful effects of acrylamide. As a result, taurine can be regarded as a potential therapeutic drug for acrylamide-induced infertility.

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Authors' Contribution

M.DZ: Data collection and interpretation, drafting and reviewing the manuscript; Z.KH: Study design, data analysis, drafting and reviewing the manuscript; MA.E: Data analysis and interpretation, drafting and reviewing the manuscript; All authors contributed to the study's conception and design and commented on the last versions of the manuscript and read and approved the final manuscript and provided an agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Conflict of Interest: None declared.

References

- Muttucumaru N, Powers SJ, Elmore JS, Dodson A, Briddon A, Mottram DS, et al. Acrylamide-forming potential of potatoes grown at different locations, and the ratio of free asparagine to reducing sugars at which free asparagine becomes a limiting factor for acrylamide formation. Food Chem. 2017;220:76-86. doi: 10.1016/j. foodchem.2016.09.199. PubMed PMID: 27855938; PubMed Central PMCID: PMCPMC5119237.
- Nan B, Yang C, Li L, Ye H, Yan H, Wang M, et al. Allicin alleviated acrylamide-induced NLRP3 inflammasome activation via oxidative stress and endoplasmic reticulum stress in Kupffer cells and SD rats liver. Food Chem Toxicol. 2021;148:111937. doi: 10.1016/j. fct.2020.111937. PubMed PMID: 33348049.
- Aldawood N, Alrezaki A, Alanazi S, Amor N, Alwasel S, Sirotkin A, et al. Acrylamide impairs ovarian function by promoting apoptosis and affecting reproductive hormone release, steroidogenesis and autophagy-related genes: An in vivo study. Ecotoxicol Environ Saf. 2020;197:110595. doi: 10.1016/j.ecoenv.2020.110595. PubMed PMID: 32304918.
- 4 Zamani E, Shokrzadeh M, Fallah M, Shaki F. A review of acrylamide toxicity and its

- mechanism. Pharmaceutical and biomedical research. 2017;3:1-7. doi: 10.18869/acad-pub.pbr.3.1.1.
- Ommati MM, Heidari R, Manthari RK, Tikka Chiranjeevi S, Niu R, Sun Z, et al. Paternal exposure to arsenic resulted in oxidative stress, autophagy, and mitochondrial impairments in the HPG axis of pubertal male offspring. Chemosphere. 2019;236:124325. doi: 10.1016/j.chemosphere.2019.07.056. PubMed PMID: 31326754.
- 6 Wang L, Ye X, Zhao T. The physiological roles of autophagy in the mammalian life cycle. Biol Rev Camb Philos Soc. 2019;94:503-16. doi: 10.1111/brv.12464. PubMed PMID: 30239126; PubMed Central PMCID: PMCPMC7379196.
- 7 Cao B, Camden AJ, Parnell LA, Mysorekar IU. Autophagy regulation of physiological and pathological processes in the female reproductive tract. Am J Reprod Immunol. 2017;77. doi: 10.1111/aji.12650. PubMed PMID: 28194822.
- Alaee S, Mirani M, Derakhshan Z, Koohpeyma F, Bakhtari A. Thymoquinone improves folliculogenesis, sexual hormones, gene expression of apoptotic markers and antioxidant enzymes in polycystic ovary syndrome rat model. Vet Med Sci. 2023;9:290-300. doi: 10.1002/vms3.958. PubMed PMID: 36104839; PubMed Central PMCID: PMCPMC9857009.
- 9 Ojatula AO. Developmental botanic: A case study on chromatographic determination of phytocompounds in the roots of anthocleista nobilis (G. Don.) as pro-fertility. Journal of Infertility and Reproductive Biology. 2021;9:7-21.
- 10 Alaee S, Khodabandeh Z, Dara M, Hosseini E, Sharma M. Curcumin mitigates acrylamide-induced ovarian antioxidant disruption and apoptosis in female Balb/c mice: A comprehensive study on gene and protein expressions. Food Sci Nutr. 2024;12:4160-72. doi: 10.1002/fsn3.4076. PubMed PMID: 38873462; PubMed Central PMCID: PMCPMC11167175.
- 11 Schaffer S, Kim HW. Effects and Mechanisms of Taurine as a Therapeutic Agent. Biomol Ther (Seoul). 2018;26:225-41. doi: 10.4062/biomolther.2017.251. PubMed PMID: 29631391; PubMed Central PMCID: PMCPMC5933890.
- 12 De Luca A, Pierno S, Camerino DC. Taurine: the appeal of a safe amino acid for skeletal muscle disorders. J Transl Med. 2015;13:243. doi: 10.1186/s12967-015-0610-1. PubMed PMID: 26208967; PubMed Central PMCID:

- PMCPMC4513970.
- 13 Baliou S, Adamaki M, Ioannou P, Pappa A, Panayiotidis MI, Spandidos DA, et al. Protective role of taurine against oxidative stress (Review). Mol Med Rep. 2021;24. doi: 10.3892/mmr.2021.12242. PubMed PMID: 34184084; PubMed Central PMCID: PMCPMC8240184.
- 14 Zhang Z, Liu D, Yi B, Liao Z, Tang L, Yin D, et al. Taurine supplementation reduces oxidative stress and protects the liver in an iron-overload murine model. Mol Med Rep. 2014;10:2255-62. doi: 10.3892/mmr.2014.2544. PubMed PMID: 25201602; PubMed Central PMCID: PMCPMC4199407.
- Mousavi K, Niknahad H, Ghalamfarsa A, Mohammadi H, Azarpira N, Ommati MM, et al. Taurine mitigates cirrhosis-associated heart injury through mitochondrial-dependent and antioxidative mechanisms. Clin Exp Hepatol. 2020;6:207-19. doi: 10.5114/ ceh.2020.99513. PubMed PMID: 33145427; PubMed Central PMCID: PMCPMC7592093.
- 16 Azami SH, Nazarian H, Abdollahifar MA, Eini F, Farsani MA, Novin MG. The antioxidant curcumin postpones ovarian aging in young and middle-aged mice. Reprod Fertil Dev. 2020;32:292-303. doi: 10.1071/RD18472. PubMed PMID: 31656219.
- 17 Golmohammadi T, Vaezi G. Effect of Curcuma Longa on In Vitro Maturation of Immature Oocytes and Embryonic Development in NMRI Mice. Journal of Mazandaran University of Medical Sciences. 2018;27:49-58.
- 18 Huynh DTM, De Tran V, Pan-On S, Pham DT. In-vivo anti-tumor activities of curcumin-solid-dispersion loaded floating tablets on stomach and lung cancers. Journal of Applied Pharmaceutical Science. 2024;14:081-7.
- 19 Chen J, Li J, Fan T, Zhong S, Qin X, Li R, et al. Protective effects of curcumin/cyclodextrin polymer inclusion complex against hydrogen peroxide-induced LO2 cells damage. Food Sci Nutr. 2022;10:1649-56. doi: 10.1002/fsn3.2787. PubMed PMID: 35592280; PubMed Central PMCID: PMCPMC9094476.
- 20 Care IoLARCo, Animals UoL. Guide for the Care and Use of Laboratory Animals. The National Academies Collection: Reports funded by National Institutes of Health. 8th ed. Institute of Laboratory Animal Resources: Washington (DC); 2011.
- 21 Daneshpazhouh H, Hayati Roodbari N, Tahamtani Y, Khodabandeh Z, Dianatpour M. Protective Effect of Docetaxel Against Autophagy-Related Genes in Vitrification of Mouse Metaphase II Oocytes. Iran J Med Sci. 2023;48:501-9. doi: 10.30476/

- IJMS.2023.88390.2811. PubMed PMID: 37786462; PubMed Central PMCID: PMCPMC10541544.
- 22 Roy D, Tomo S, Modi A, Purohit P, Sharma P. Optimising total RNA quality and quantity by phenol-chloroform extraction method from human visceral adipose tissue: A standardisation study. MethodsX. 2020;7:101113. doi: 10.1016/j.mex.2020.101113. PubMed PMID: 33204654; PubMed Central PMCID: PMCPMC7653057.
- 23 Rao X, Huang X, Zhou Z, Lin X. An improvement of the 2[^](-delta delta CT) method for quantitative real-time polymerase chain reaction data analysis. Biostat Bioinforma Biomath. 2013;3:71-85. PubMed PMID: 25558171; PubMed Central PMCID: PMCPMC4280562.
- 24 Ommati MM, Heidari R, Jamshidzadeh A, Zamiri MJ, Sun Z, Sabouri S, et al. Dual effects of sulfasalazine on rat sperm characteristics, spermatogenesis, and steroidogenesis in two experimental models. Toxicol Lett. 2018;284:46-55. doi: 10.1016/j. toxlet.2017.11.034. PubMed PMID: 29197623.
- 25 Li S, Zhao H, Wang Y, Shao Y, Wang B, Wang Y, et al. Regulation of autophagy factors by oxidative stress and cardiac enzymes imbalance during arsenic or/and copper induced cardiotoxicity in Gallus gallus. Ecotoxicol Environ Saf. 2018;148:125-34. doi: 10.1016/j.ecoenv.2017.10.018. PubMed PMID: 29035754.
- 26 Song D, Xu C, Holck AL, Liu R. Acrylamide inhibits autophagy, induces apoptosis and alters cellular metabolic profiles. Ecotoxicol Environ Saf. 2021;208:111543. doi: 10.1016/j.ecoenv.2020.111543. PubMed PMID: 33396091.
- 27 Park H, Cho M, Do Y, Park JK, Bae SJ, Joo J, et al. Autophagy as a Therapeutic Target of Natural Products Enhancing Embryo Implantation. Pharmaceuticals (Basel). 2021;15. doi: 10.3390/ph15010053. PubMed PMID: 35056110; PubMed Central PMCID: PMCPMC8779555.
- 28 Zheng Y, Ma L, Liu N, Tang X, Guo S, Zhang B, et al. Autophagy and Apoptosis of Porcine Ovarian Granulosa Cells During Follicular Development. Animals (Basel). 2019;9. doi: 10.3390/ani9121111. PubMed PMID: 31835576; PubMed Central PMCID: PMCPMC6940823.
- 29 Saifi B, Haftcheshmeh SM, Feligioni M, Izadpanah E, Rahimi K, Hassanzadeh K, et al. An overview of the therapeutic effects of curcumin in reproductive disorders with a focus

- on the antiinflammatory and immunomodulatory activities. Phytother Res. 2022;36:808-23. doi: 10.1002/ptr.7360. PubMed PMID: 35041229.
- 30 Maiti P, Scott J, Sengupta D, Al-Gharaibeh A, Dunbar GL. Curcumin and Solid Lipid Curcumin Particles Induce Autophagy, but Inhibit Mitophagy and the PI3K-Akt/mTOR Pathway in Cultured Glioblastoma Cells. Int J Mol Sci. 2019;20. doi: 10.3390/ijms20020399. PubMed PMID: 30669284; PubMed Central PMCID: PMCPMC6359162.
- 31 Peters AE, Mihalas BP, Bromfield EG, Roman SD, Nixon B, Sutherland JM. Autophagy in Female Fertility: A Role in Oxidative Stress and Aging. Antioxid Redox Signal. 2020;32:550-68. doi: 10.1089/ars.2019.7986. PubMed PMID: 31892284.
- 32 Khayatan D, Razavi SM, Arab ZN, Hosseini Y, Niknejad A, Momtaz S, et al. Superoxide dismutase: a key target for the neuroprotective effects of curcumin. Mol Cell Biochem. 2024;479:693-705. doi: 10.1007/s11010-023-04757-5. PubMed PMID: 37166541.
- 33 Gao N, Yao X, Jiang L, Yang L, Qiu T, Wang Z, et al. Taurine improves low-level inorganic arsenic-induced insulin resistance by activating PPARgamma-mTORC2 signalling and inhibiting hepatic autophagy. J Cell Physiol. 2019;234:5143-52. doi: 10.1002/jcp.27318. PubMed PMID: 30362509.
- 34 Kaneko H, Kobayashi M, Mizunoe Y, Yoshida M, Yasukawa H, Hoshino S, et al. Taurine is an amino acid with the ability to activate autophagy in adipocytes. Amino Acids. 2018;50:527-35. doi: 10.1007/s00726-018-2550-6. PubMed PMID: 29523960.
- 35 Liu K, Mao X, Li H, Liu D, Le G, Gan F, et al.

- Regulation of taurine in OTA-induced apoptosis and autophagy. Toxicon. 2020;181:82-90. doi: 10.1016/j.toxicon.2020.04.097. PubMed PMID: 32371067.
- 36 Sun Y, Dai S, Tao J, Li Y, He Z, Liu Q, et al. Taurine suppresses ROS-dependent autophagy via activating Akt/mTOR signaling pathway in calcium oxalate crystals-induced renal tubular epithelial cell injury. Aging (Albany NY). 2020;12:17353-66. doi: 10.18632/aging.103730. PubMed PMID: 32931452; PubMed Central PMCID: PMCPMC7521519.
- 37 Tan X, Ye J, Liu W, Zhao B, Shi X, Zhang C, et al. Acrylamide aggravates cognitive deficits at night period via the gut-brain axis by reprogramming the brain circadian clock. Arch Toxicol. 2019;93:467-86. doi: 10.1007/s00204-018-2340-7. PubMed PMID: 30374679.
- 38 Schaaf MB, Keulers TG, Vooijs MA, Rouschop KM. LC3/GABARAP family proteins: autophagy-(un)related functions. FASEB J. 2016;30:3961-78. doi: 10.1096/fj.201600698R. PubMed PMID: 27601442.
- 39 Yang N, Li L, Li Z, Ni C, Cao Y, Liu T, et al. Protective effect of dapsone on cognitive impairment induced by propofol involves hippocampal autophagy. Neurosci Lett. 2017;649:85-92. doi: 10.1016/j.neulet.2017.04.019. PubMed PMID: 28411068.
- 40 Zou H, Wang T, Yuan J, Sun J, Yuan Y, Gu J, et al. Cadmium-induced cytotoxicity in mouse liver cells is associated with the disruption of autophagic flux via inhibiting the fusion of autophagosomes and lysosomes. Toxicol Lett. 2020;321:32-43. doi: 10.1016/j.toxlet.2019.12.019. PubMed PMID: 31862506.